

# Utilizing Mass-Directed SFC in Support of Medicinal Chemistry

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# Overview

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- Introduction of Technology Enabled Synthesis group (TES) and associated Purification Group
- Highlight deficiencies of reversed-phase, preparative HPLC currently in use
- Why evaluate SFC to overcome these deficiencies
- How SFC platform has been incorporated in high-throughput area
- Summarize results



# Limitations of Current Reversed-Phase HPLC Platform

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- Approaching optimal cycle time of prep LC/MS while retaining respectable resolution.
  - 25 mL/min on a 2 cm i.d. column
- Drying aqueous based HPLC fractions is very slow.
  - At least 8 hours in high speed rotary vacuum evaporation system.

**THE ANSWER: Mass-Directed SFC?**



# Why SFC???

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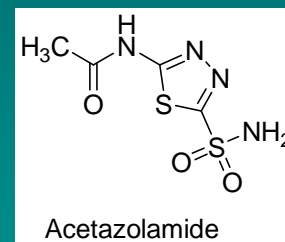
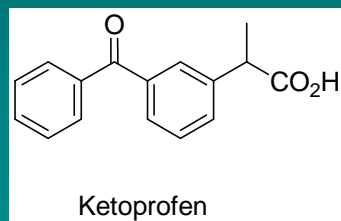
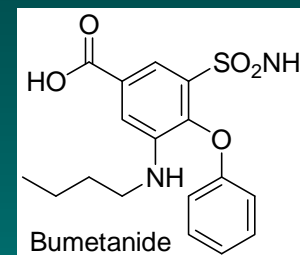
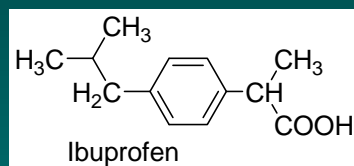
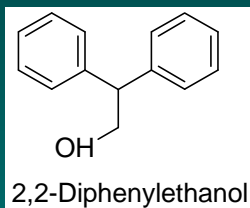
- More efficient separation per unit time compared to HPLC (3-4 times)
- Complimentary separation technique providing varied selectivity
- Very limited modifier use for reduced salt forms compared to HPLC
- A “Green Technology”
- Dramatically reduce dry down time compared to aqueous based HPLC fractions (30mL aqueous to 3mL MeOH)



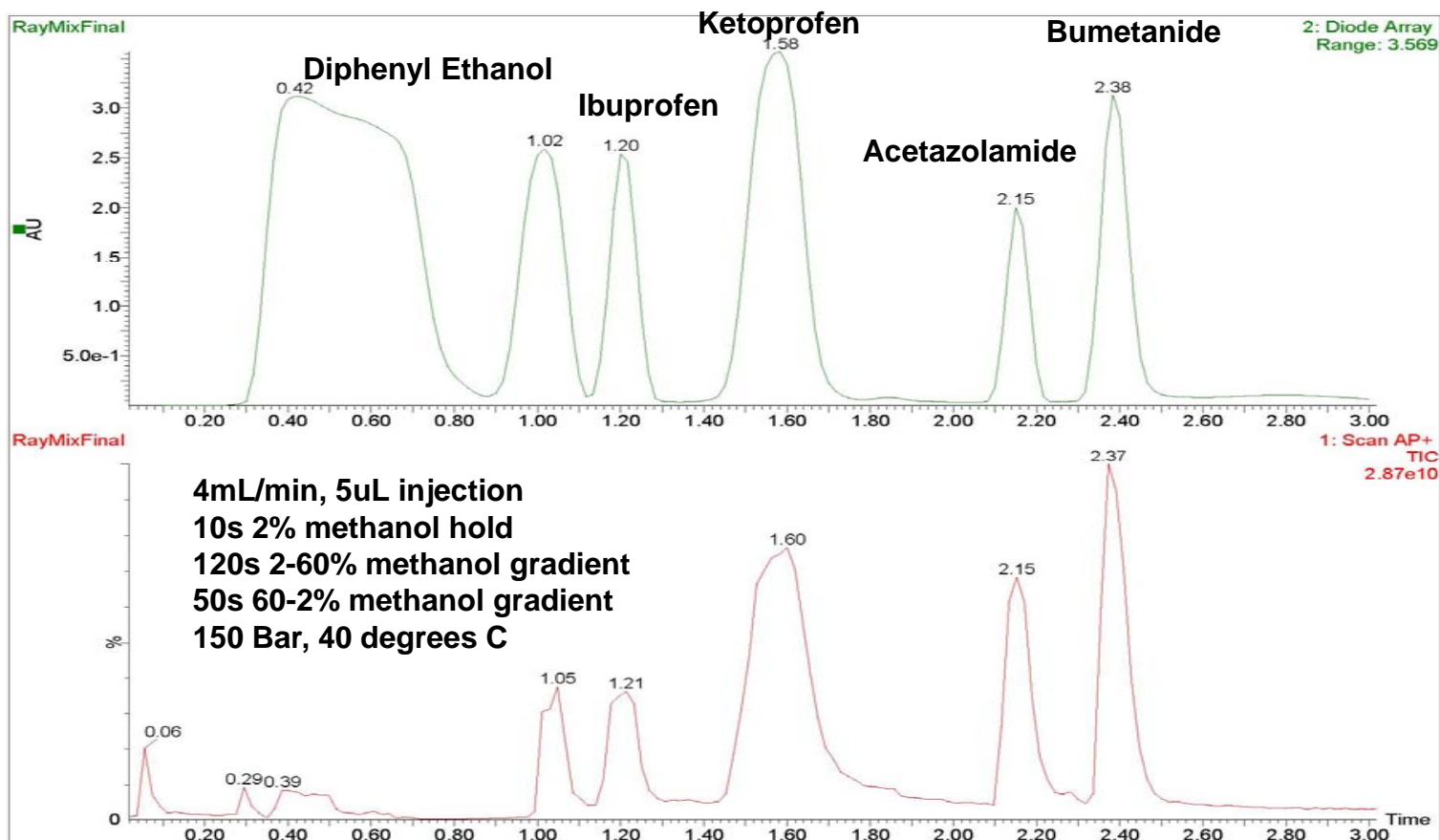
# Analytical SFC/MS



# Structures Of Test Components



# Analytical SFC/MS Screen



# SFC MS Prep-30



# Benefit of Focused Gradient Prep

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- Focused gradients
  - Don't waste time evenly spacing all peaks with the entire gradient range (5-50)
  - Focus the gradient range around desired compound
    - Combines and elutes weaker retained species immediately after void
    - Maximizes resolution resulting in accurate fractionation of single (or multiple) peak(s) of interest
    - Combines and elutes stronger retained species during the 100% organic blow-off period



# Narrow Gradient Table in FractionLynx

autopurify\_1.flp - AutoPurify

File Edit View Help

Walk-up | MS+ Data | MS- Data | DAD Data | Instrument | MS Process | Spectrum Test | Printing | Chromatogram Test  
Purification Strategy | Analytical Interpretation | Generic Method | Narrow Method | File Creation Options | Automatic Stages

Narrow Method Settings

The Narrow Method is selected using the Retention Time of the peak from the Analytical Run.

Method	Start RT	End RT	Inlet Method	Switch Method	Pre Run Method	MIT File
NarrowA	0.00	1.15	2_5_9min	-None-	prerun2	aMIT_2_5
NarrowB	1.15	1.35	5_10_9min	-None-	prerun5	aMIT_5_10
NarrowC	1.35	1.55	7_12_9min	-None-	prerun7	aMIT_7_12
NarrowD	1.55	1.75	10_15_9min	-None-	prerun10	aMIT_10_15
NarrowE	1.75	1.90	12_18_9min	-None-	prerun12	aMIT_12_18
NarrowF	1.90	2.10	15_20_9min	-None-	prerun15	aMIT_15_20

Common Settings for All Narrow Methods

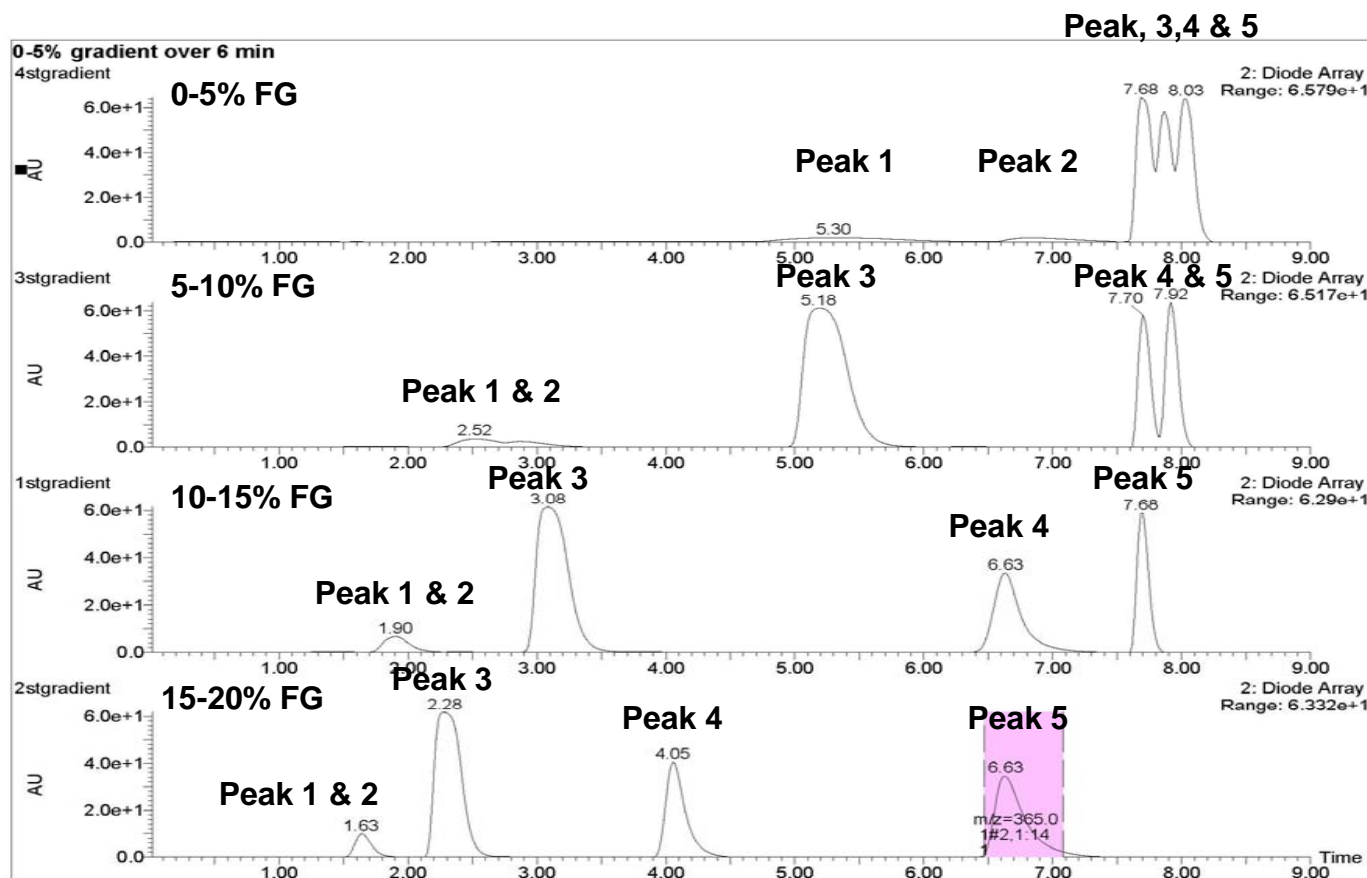
Method	MS Method	Tune File	Fraction File
Settings	9min_MS_ctc	libtune	Ray_libfrac

Peak Lies in OverLap between Narrow Methods

Use early running method     Use late running method



# Std Mix on Focused Gradients

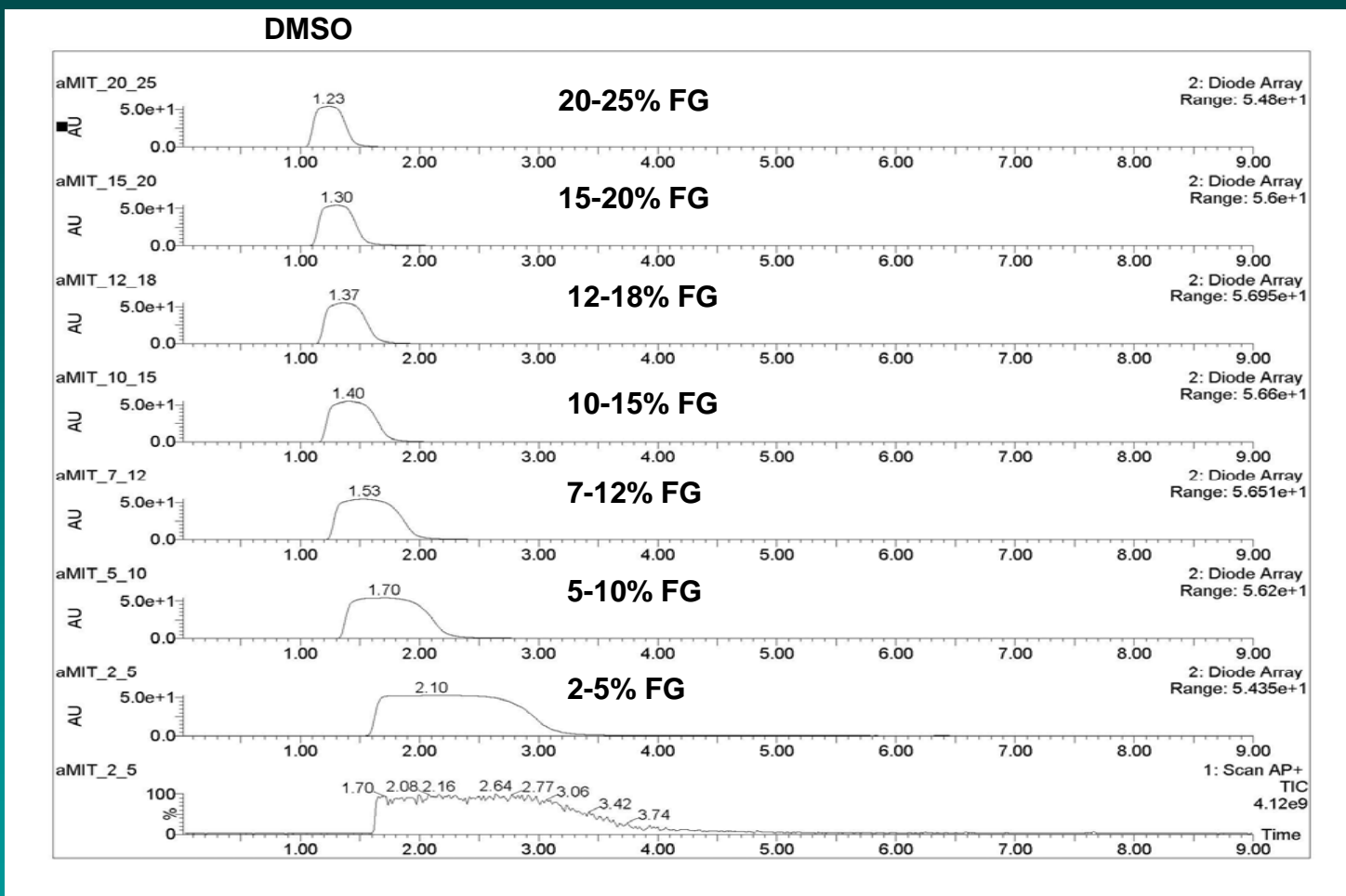


1cm i.d. x 10cm, 5 $\mu$ m, Ethylpyridine  
20mL/min, 120 Bar, 40 degrees C

6 min focused gradient  
1 min 60% flush  
2 min 60-2% reverse gradient



# Modifier Stream Injection/Loop Clearance Time



1cm i.d. x 10cm, 5 $\mu$ m, Ethylpyridine  
20mL/min, 120 Bar, 40 degrees C

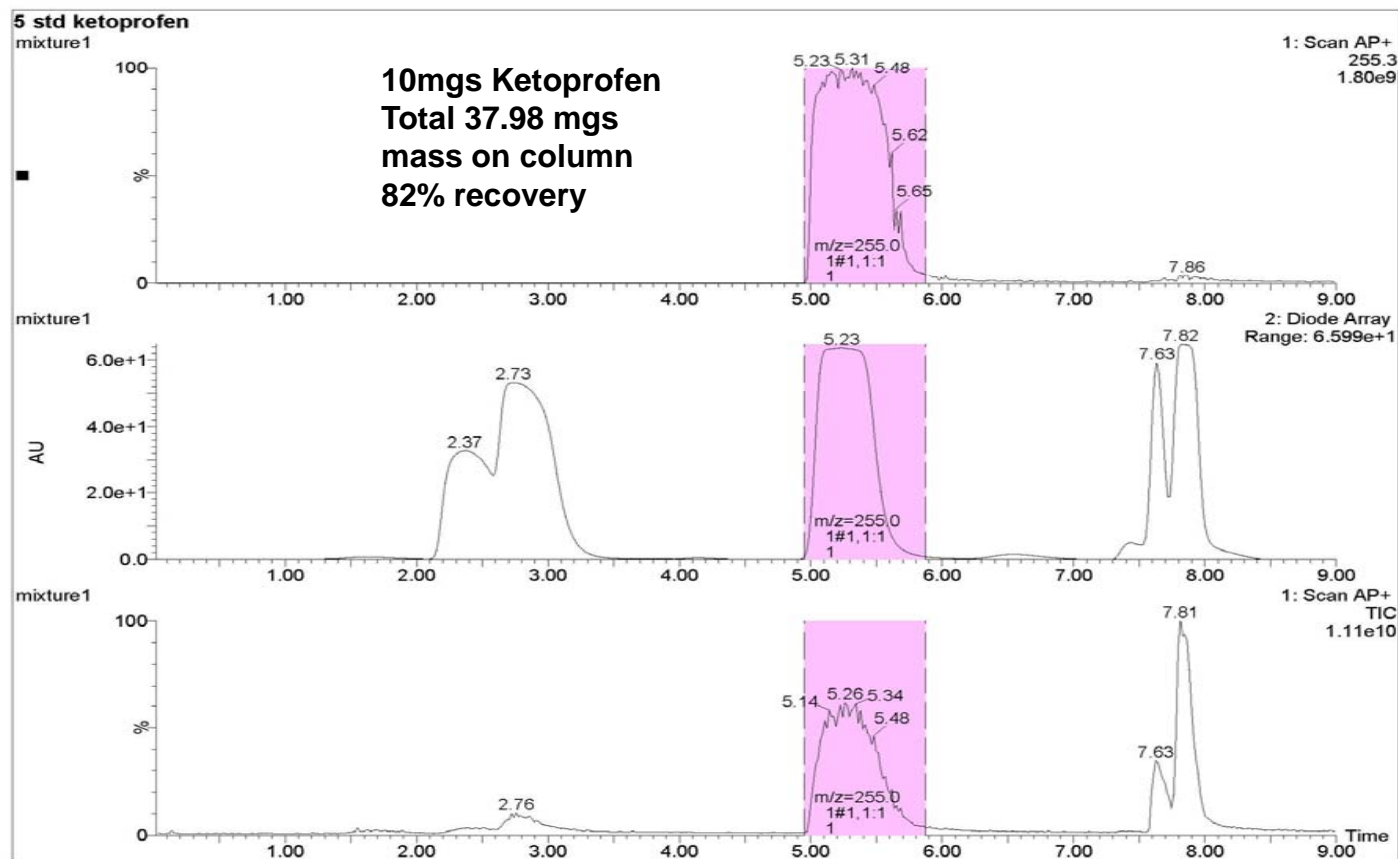
6 min focused gradient

1 min 60% flush

2 min 60-2% reverse gradient



# Single Peak Fractionation

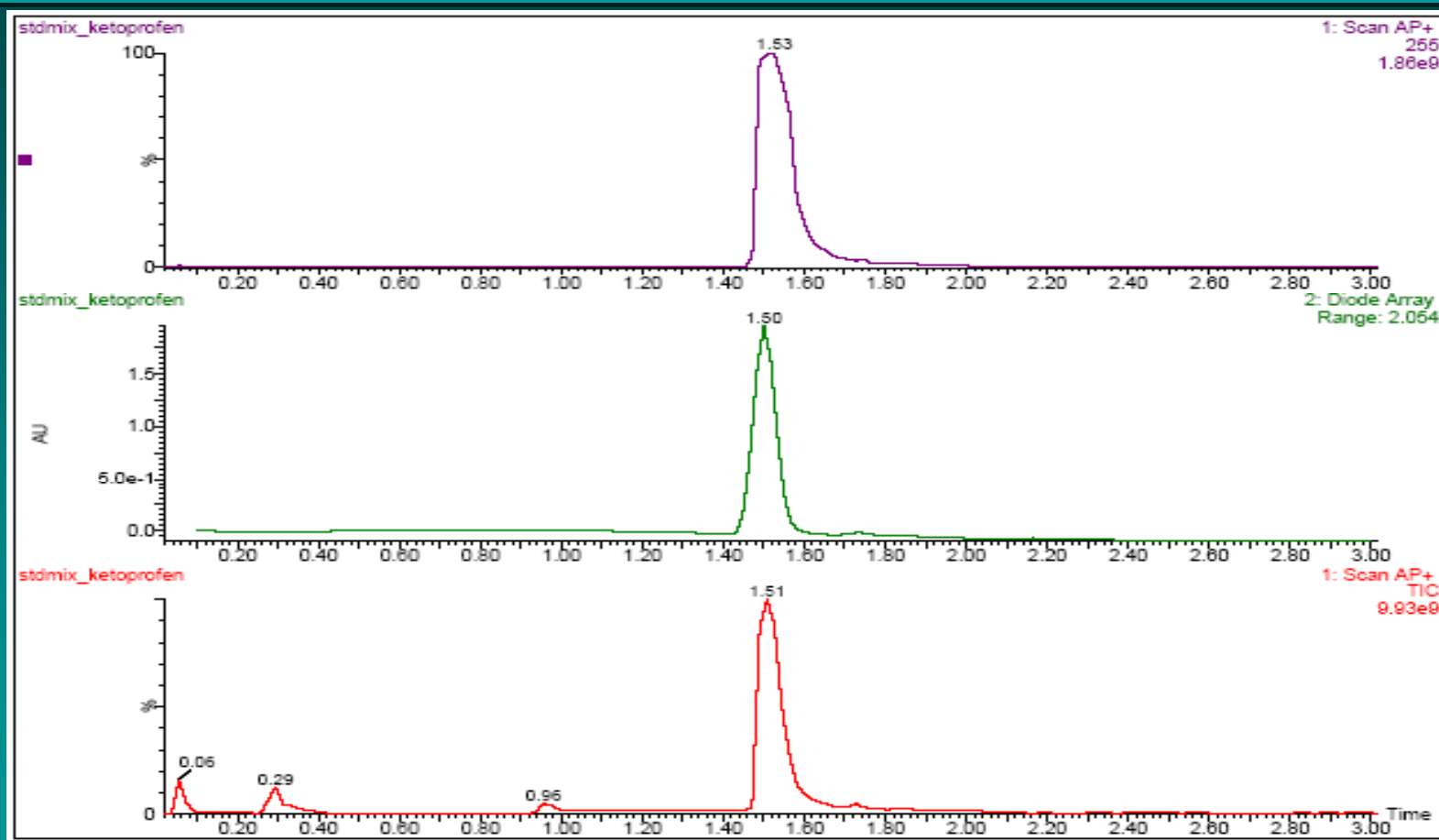


1cm i.d. x 10cm, 5 $\mu$ m, Ethylpyridine  
20mL/min, 120 Bar, 40 degrees C

6 min 5-10% methanol  
1 min 60% flush  
2 min 60-2% reverse gradient



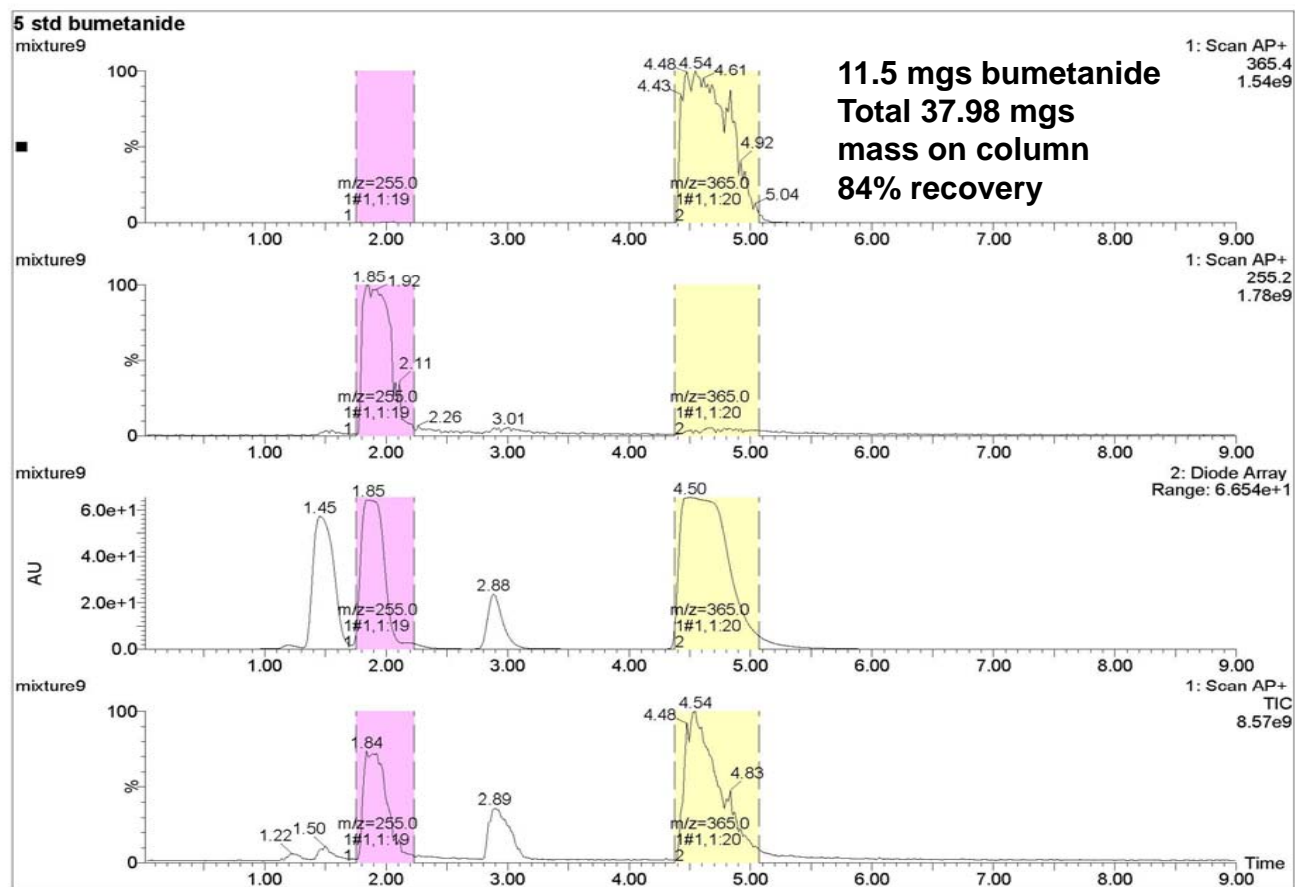
# 100% Purity of Ketoprofen



50mm x 4.6mm, 5 $\mu$ m, Ethylpyridine Column  
4mL/min  
10s 2% methanol hold  
120s 2-60% methanol gradient  
50s 60-2% methanol gradient



# Multiple Peak Fractionation

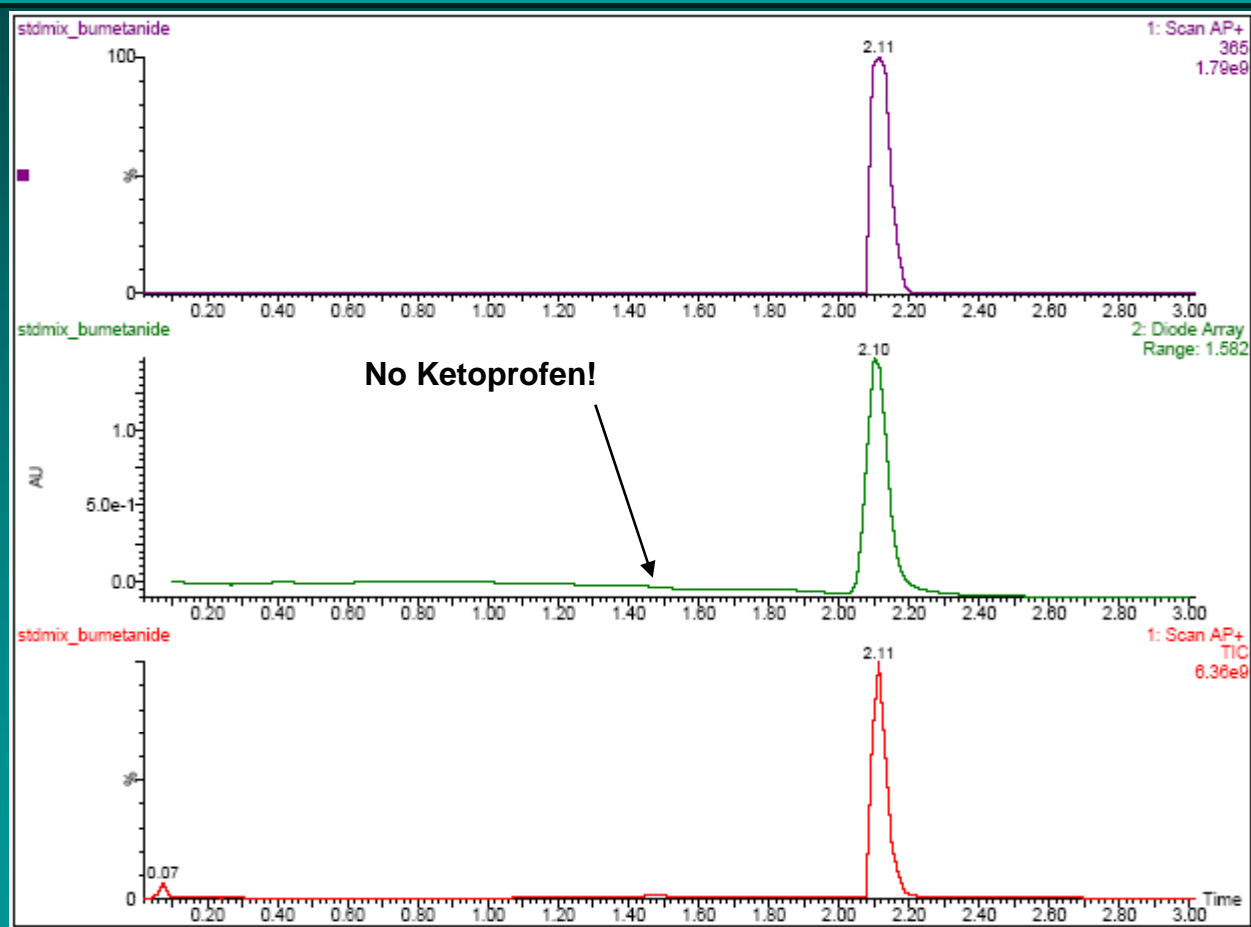


1cm i.d. x 10cm, 5um, Ethylpyridine  
20mL/min, 120 Bar, 40 degrees C

6 min 20-25% methanol  
1 min 60% flush  
2 min 60-2% reverse gradient



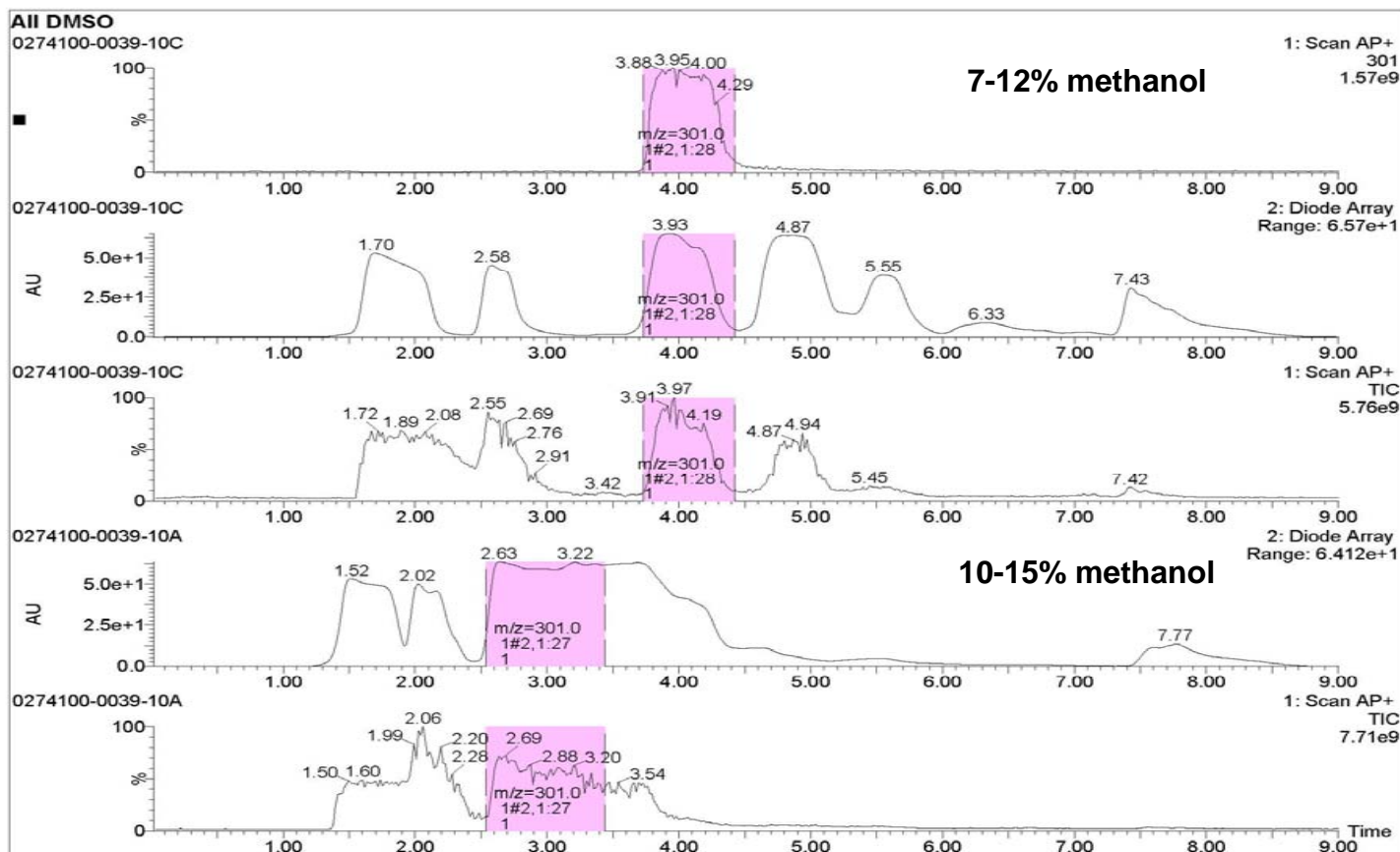
# 100% Purity, No Carry-Over



50mm x 4.6mm, 5µm, Ethylpyridine Column  
4mL/min  
10s 2% methanol hold  
120s 2-60% methanol gradient  
50s 60-2% methanol gradient



# Fine Tuning Focused Gradients



1cm i.d. x 10cm, 5 $\mu$ m, Ethylpyridine  
20mL/min, 120 Bar, 40 degrees C

6 min focused gradient  
1 min 60% flush  
2 min 60-2% reverse gradient



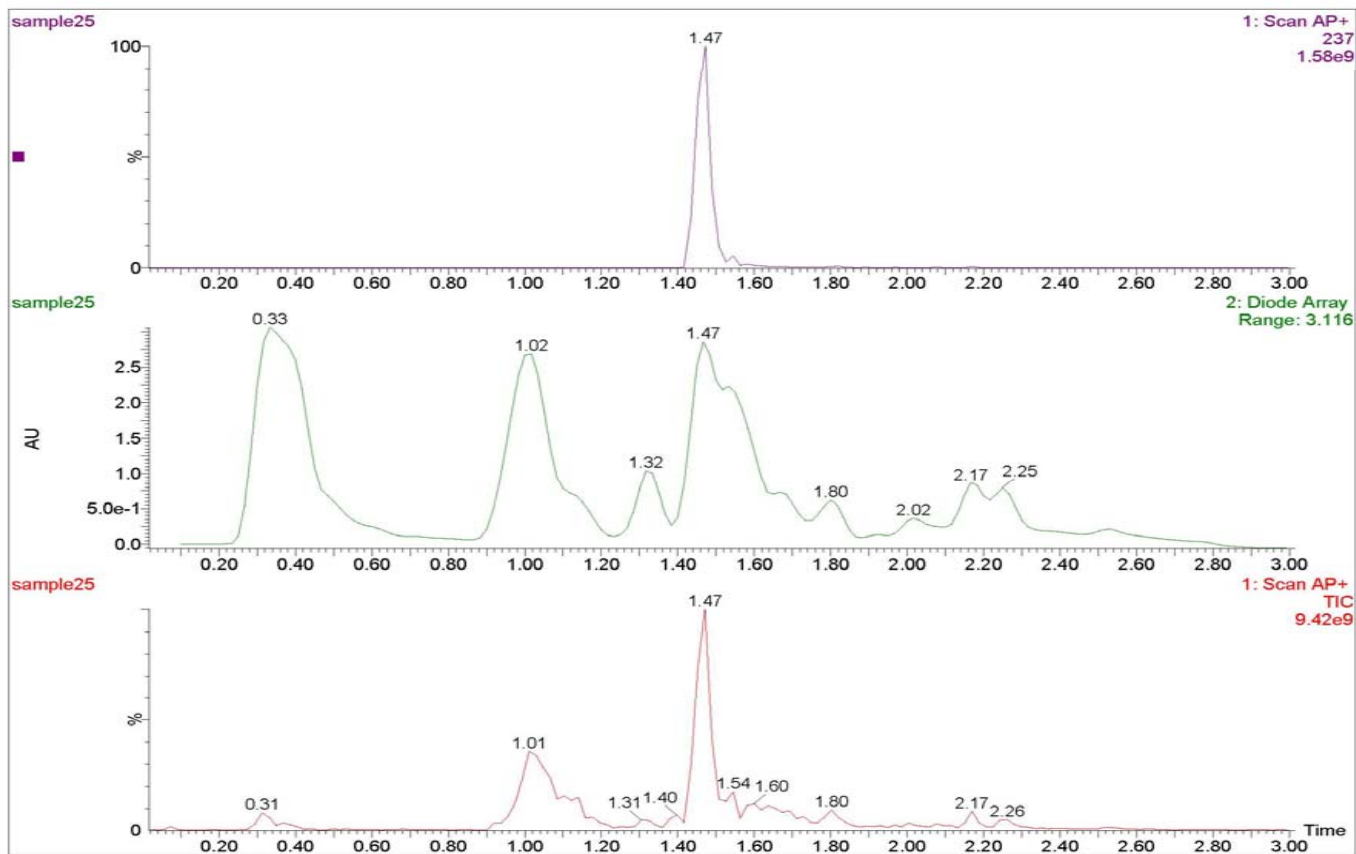
# Status Update

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- Comparable recovery to RP Prep HPLC, mid eighty percentages
- No carryover between fractions
- Can inject 1/3 of crude product, 333uL, without mass overloading column
- DMSO does not exhibit detrimental strong solvent effect
- Mapping of analytical retention times to focused gradient complete, and automated
- Ready for automated purification!!!! **ROLL THE SAMPLES**



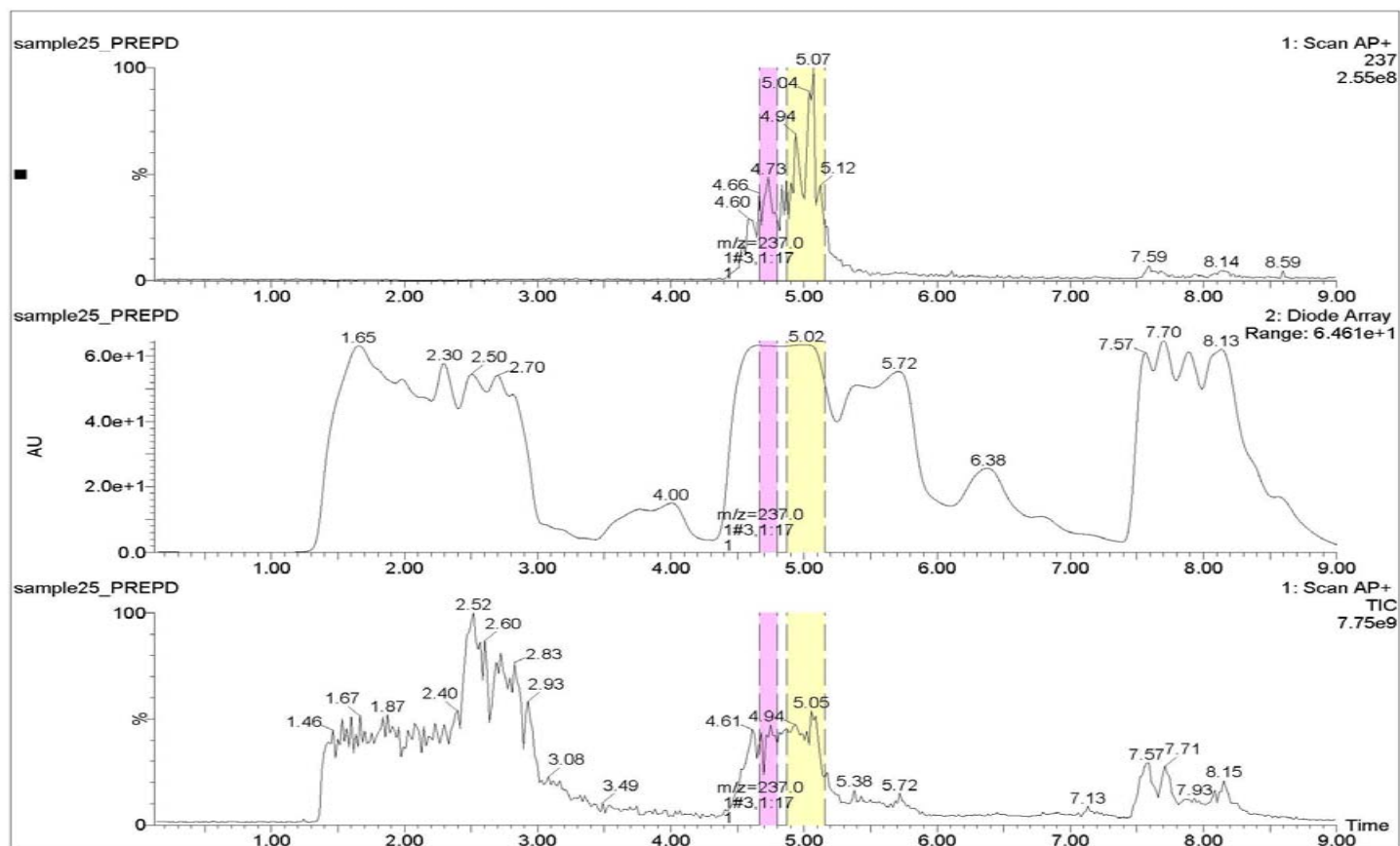
# AutoPurify: Analytical Sample One



50mm x 4.6mm, 5 $\mu$ m, Ethylpyridine Column  
4mL/min  
10s 2% methanol hold  
120s 2-60% methanol gradient  
50s 60-2% methanol gradient



# AutoPurify: Prep Sample One

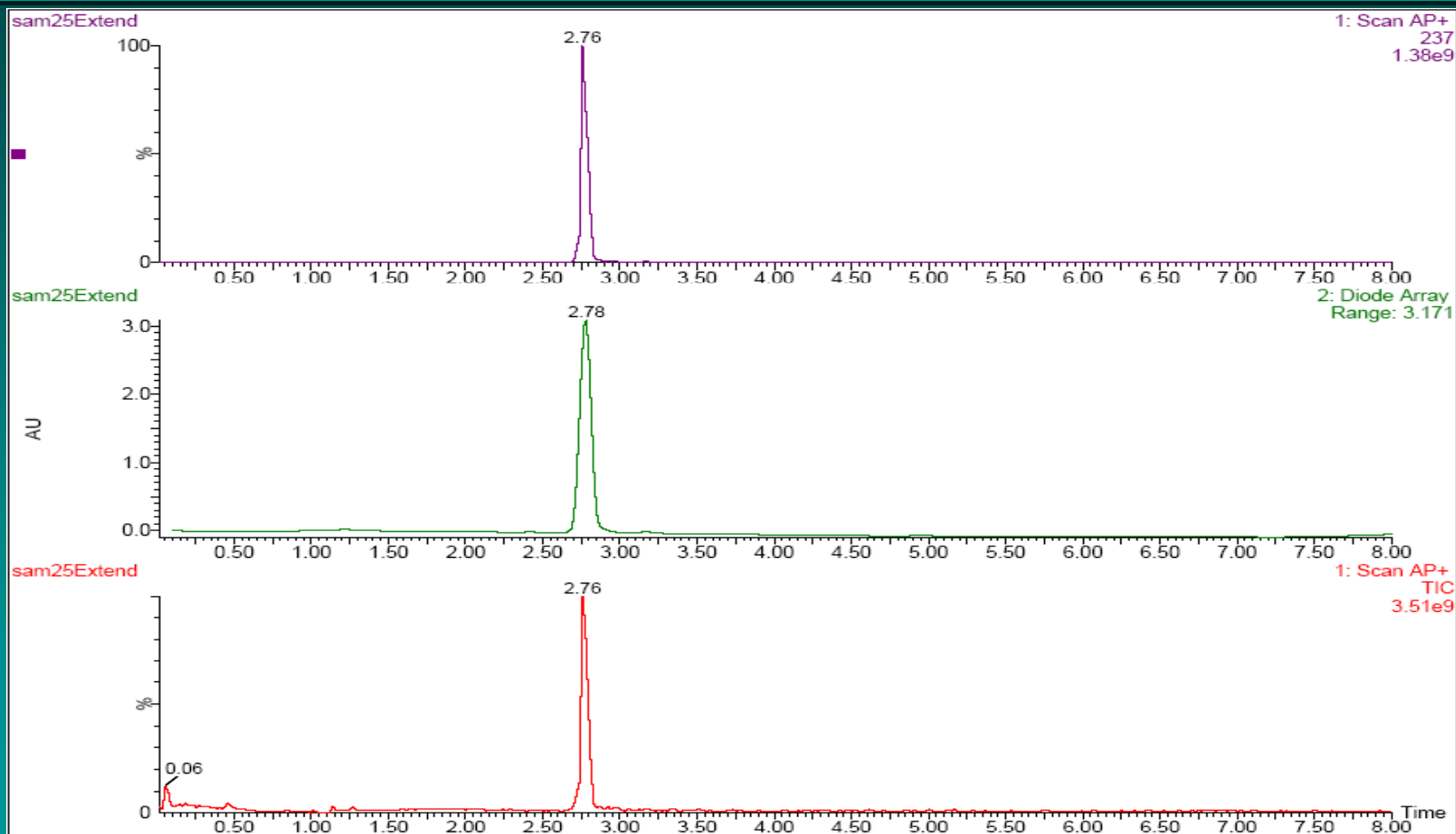


1cm i.d. x 10cm, 5 $\mu$ m, Ethylpyridine  
 20mL/min, 120 Bar, 40 degrees C  
 333 $\mu$ L injection volume

6 min 7-12% methanol  
 1 min 60% flush  
 2 min 60-2% reverse gradient



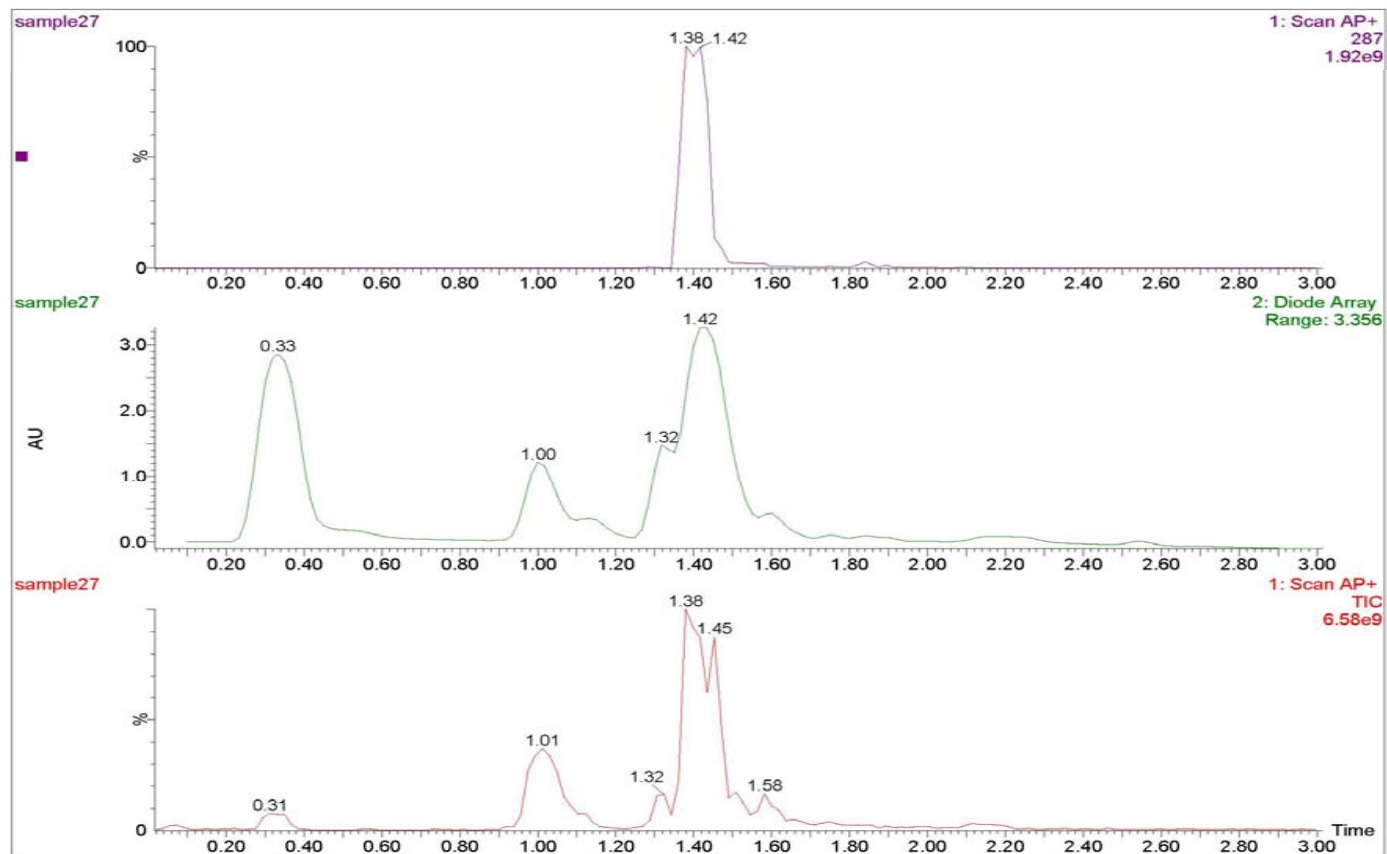
# AutoPurify: Pure Sample One



100mm x 4.6mm, 5 $\mu$ m, Ethylpyridine Column  
4mL/min  
360s 2-60% methanol gradient  
120s 60-2% methanol gradient



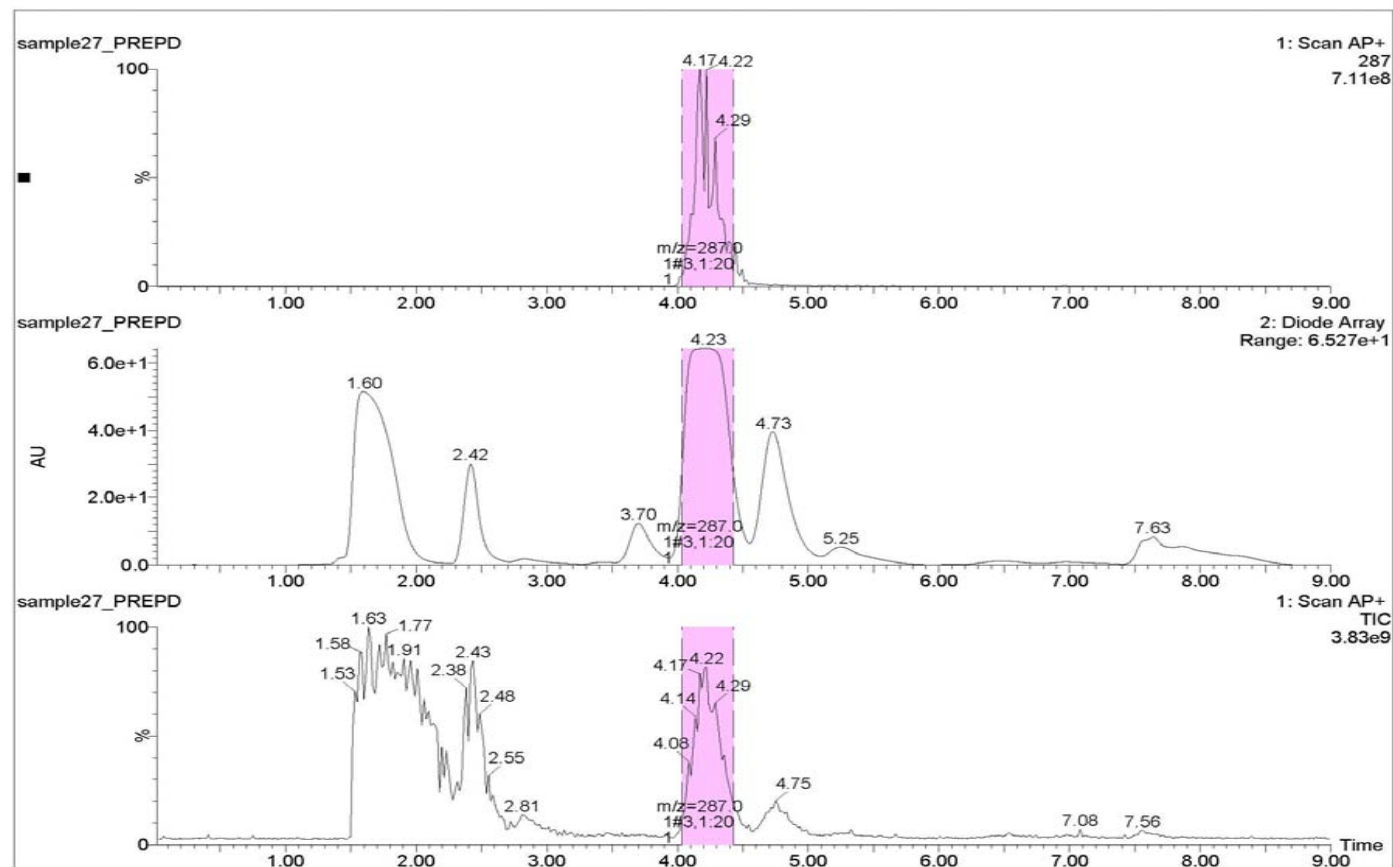
# AutoPurify: Analytical Sample Two



50mm x 4.6mm, 5 $\mu$ m, Ethylpyridine Column  
4mL/min  
10s 2% methanol hold  
120s 2-60% methanol gradient  
50s 60-2% methanol gradient



# AutoPurify: Prep Sample Two

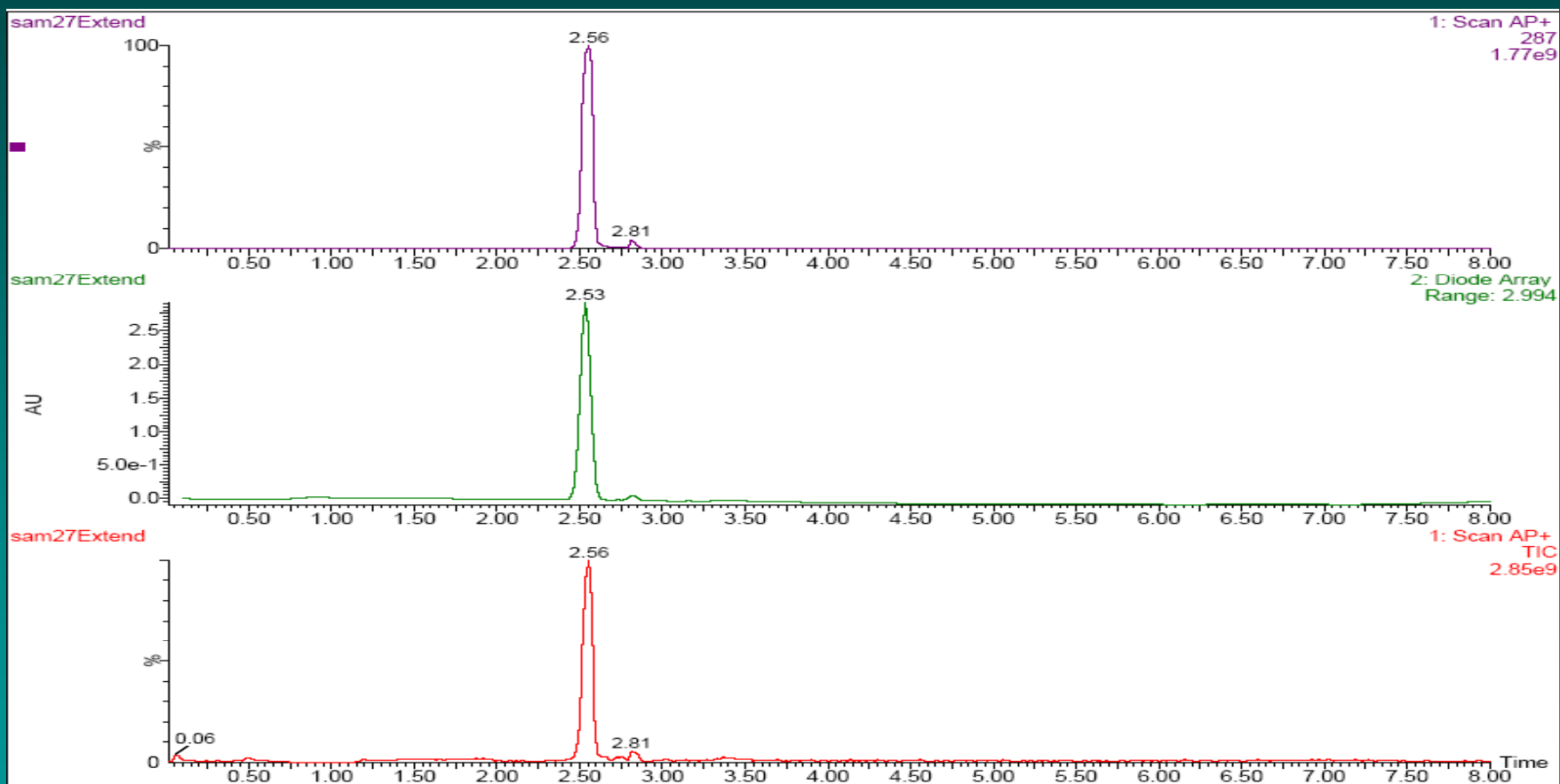


1cm i.d. x 10cm, 5 $\mu$ m, Ethylpyridine  
20mL/min, 120 Bar, 40 degrees C  
333 $\mu$ L injection volume

6 min 7-12% methanol  
1 min 60% flush  
2 min 60-2% reverse gradient



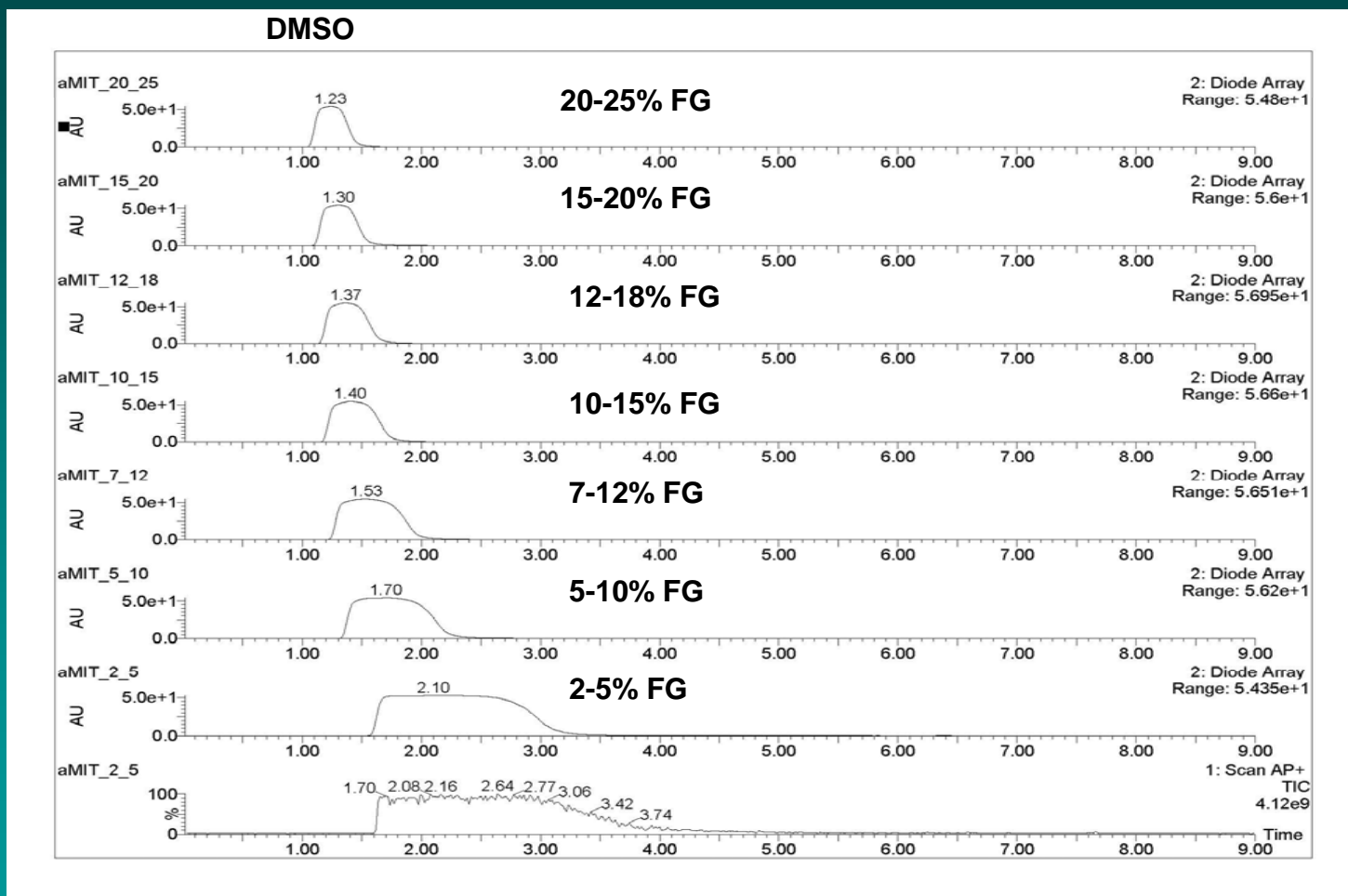
# AutoPurify: Pure Sample Two



100mm x 4.6mm, 5um, Ethylpyridine Column  
4mL/min  
360s 2-60% methanol gradient  
120s 60-2% methanol gradient



# Variable Timing of Modifier Injection



1cm i.d. x 10cm, 5 $\mu$ m, Ethylpyridine  
20mL/min, 120 Bar, 40 degrees C

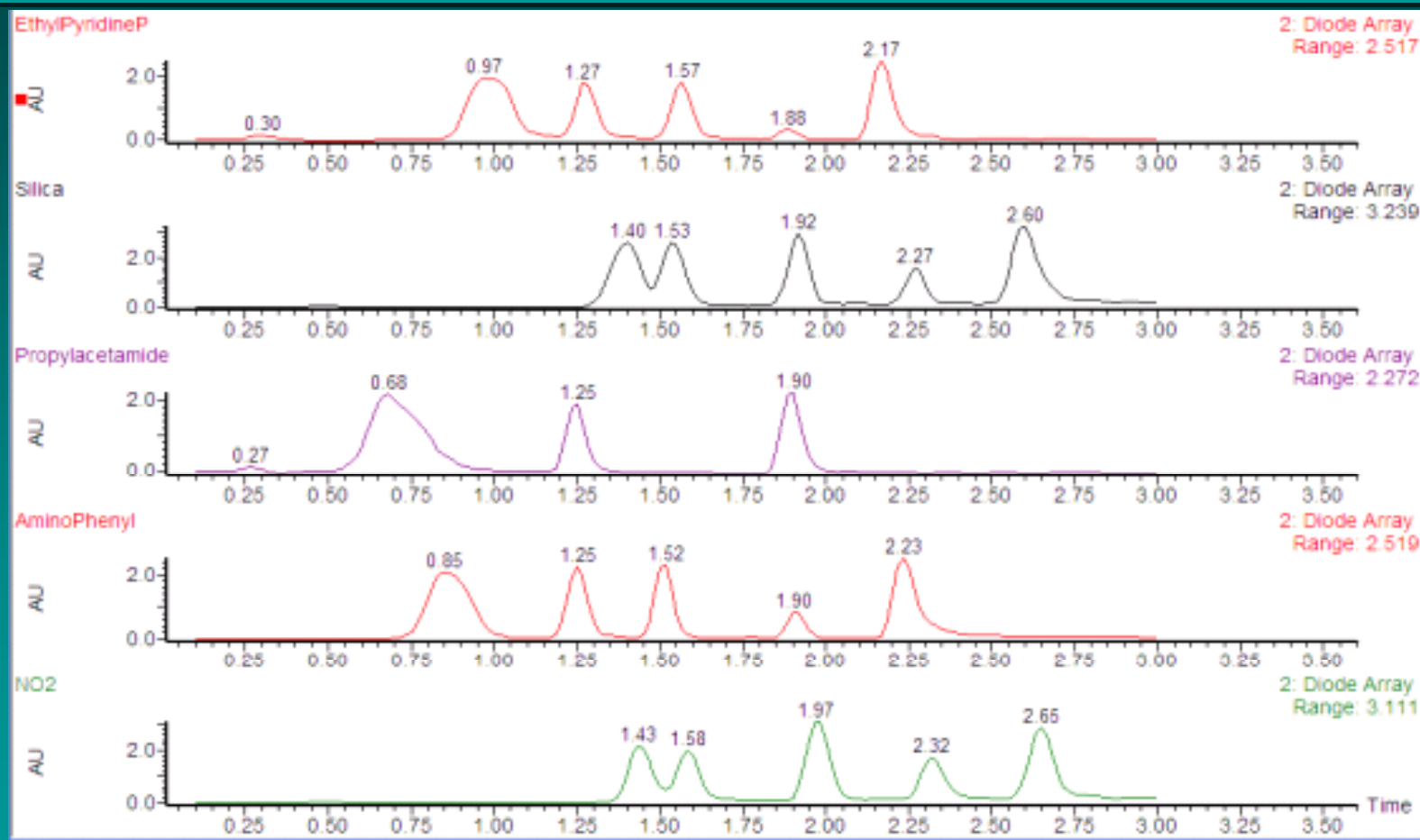
6 min focused gradient

1 min 60% flush

2 min 60-2% reverse gradient



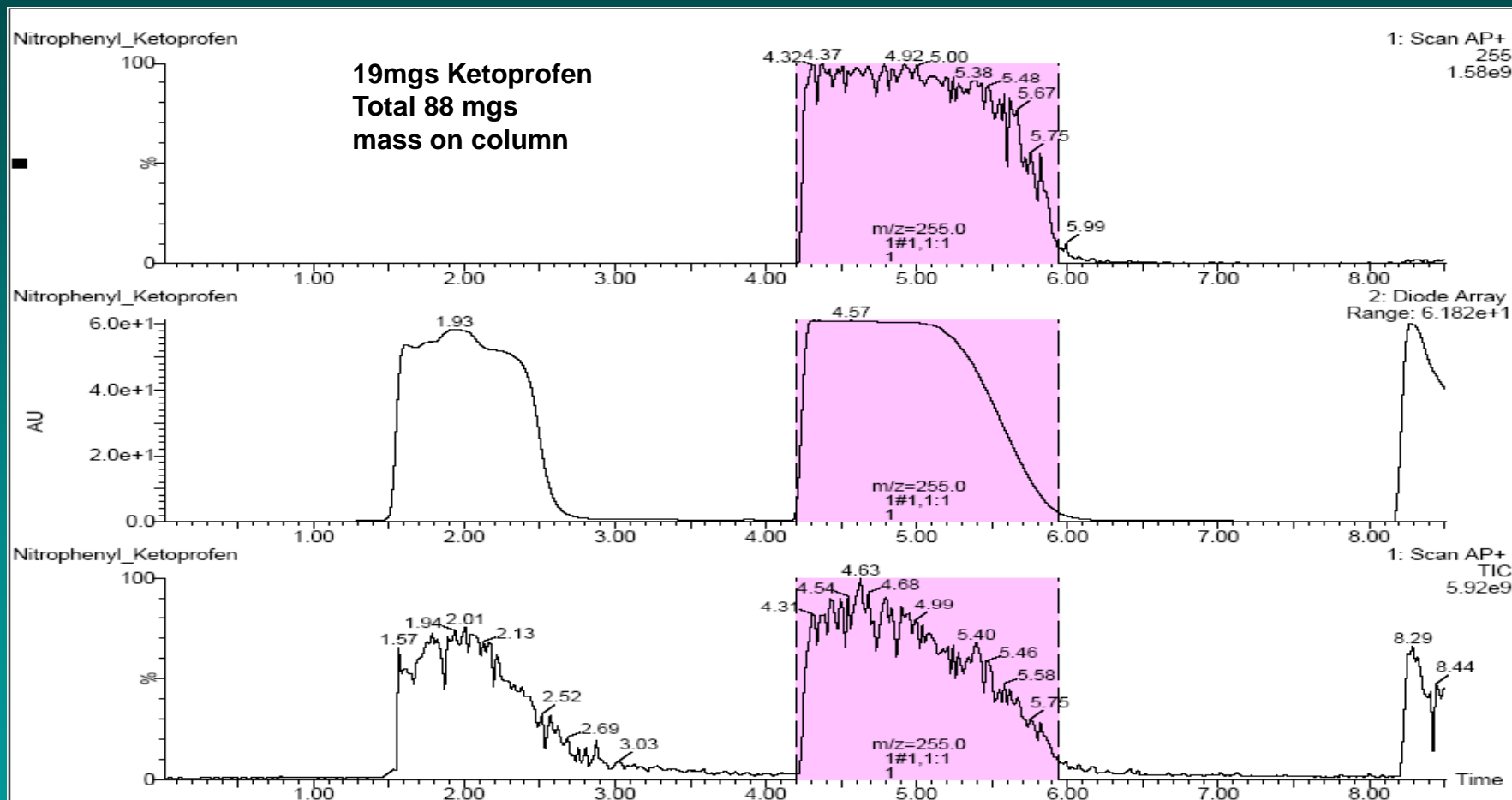
# Changing Selectivity / Different Columns



50mm x 4.6mm, 5µm, Stationary phase inside.  
4mL/min  
10s 2% methanol hold  
120s 2-60% methanol gradient  
50s 60-2% methanol gradient



# Standard Comparison: NO<sub>2</sub> Versus EP

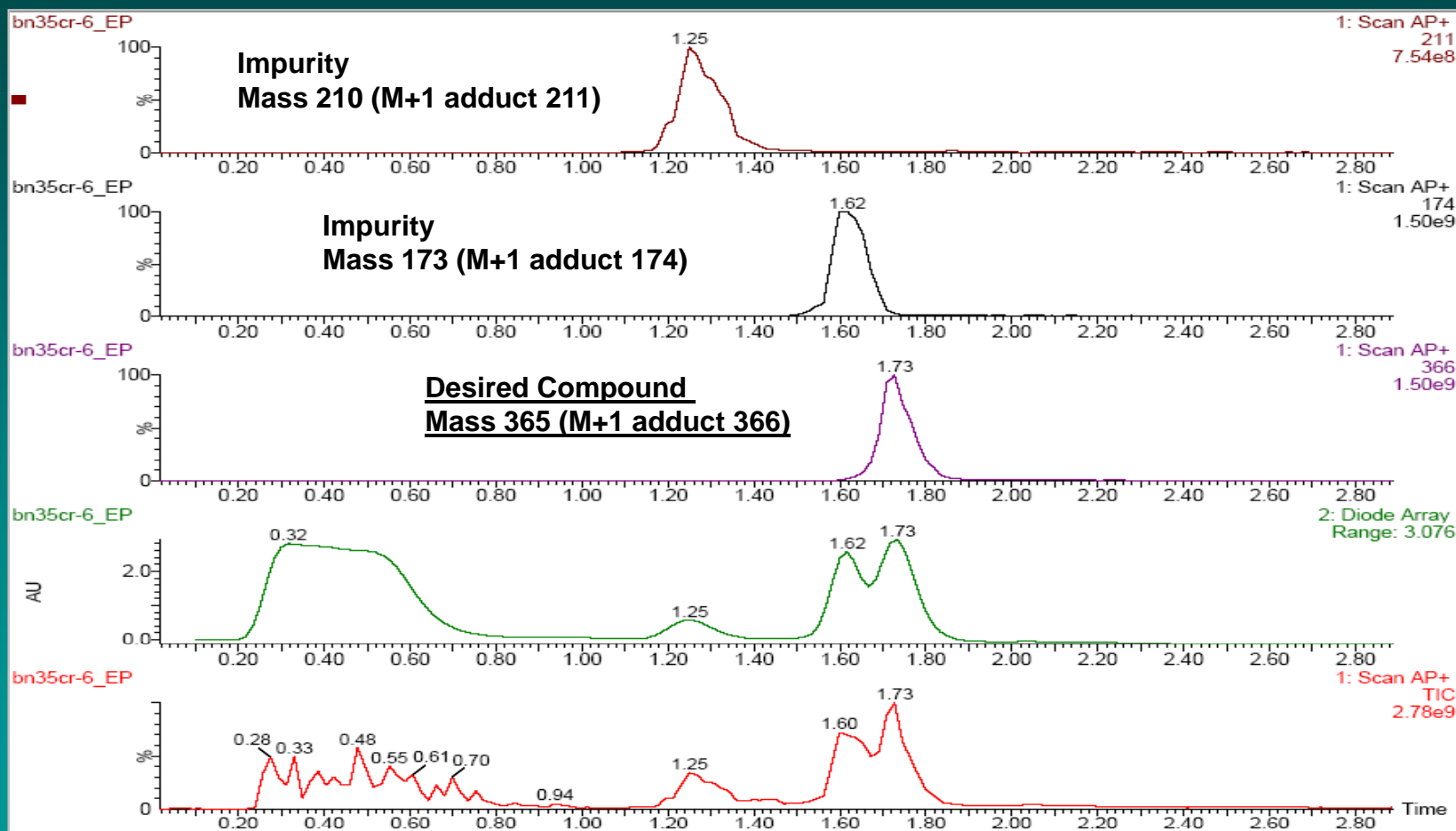


1cm i.d. x 10cm, 5 $\mu$ m, Nitrophenyl  
20mL/min, 120 Bar, 40 degrees C  
350uL injection volume

6 min 15-20% methanol  
1 min 60% flush  
2 min 60-2% reverse gradient



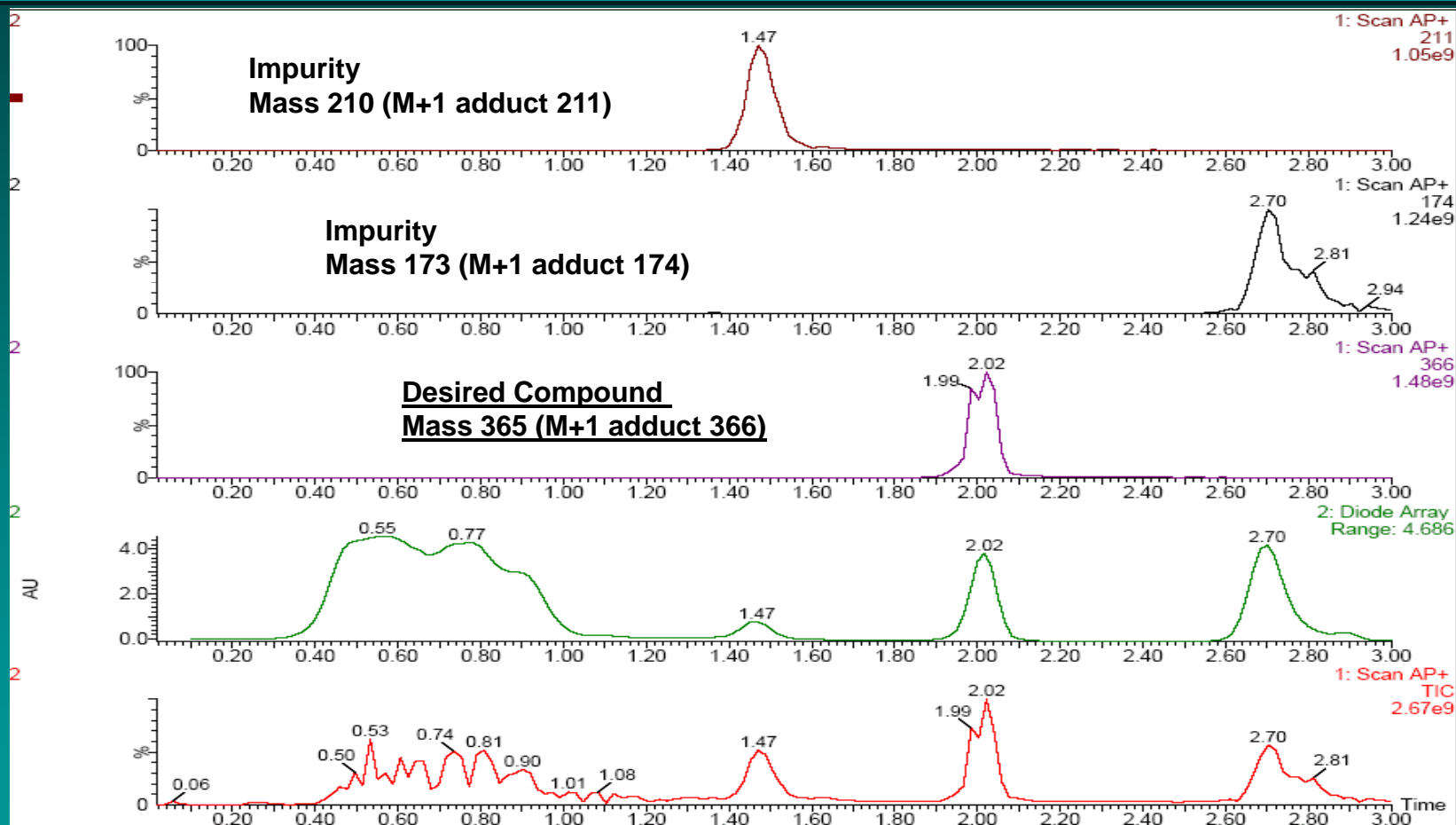
# Sample One Comparison: NO2 Versus EP



50mm x 4.6mm, 5um, Ethylpyridine Column  
4mL/min  
10s 2% methanol hold  
120s 2-60% methanol gradient  
50s 60-2% methanol gradient



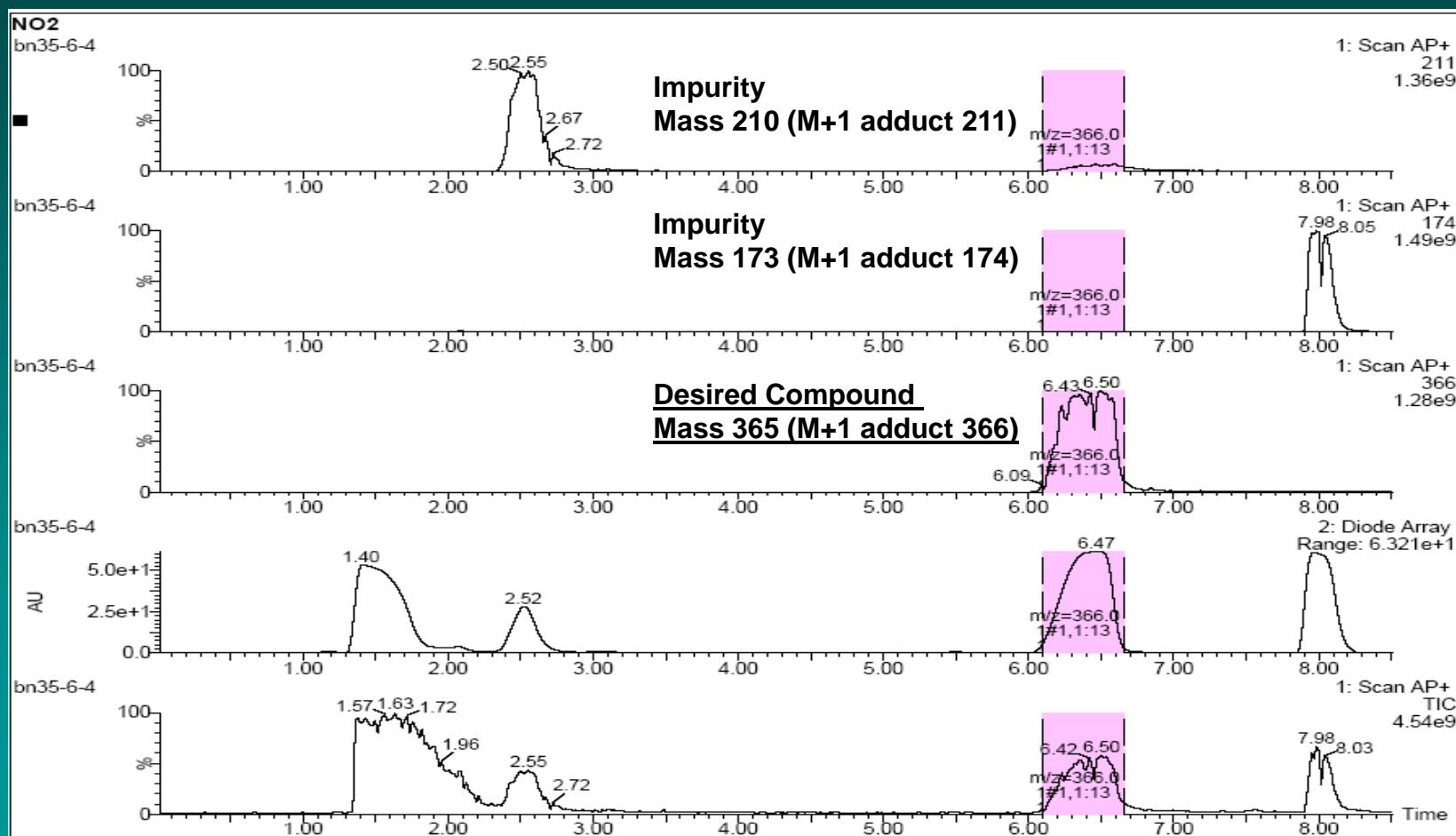
# Sample One Comparison: NO2 Versus EP



50mm x 4.6mm, 5um, Nitrophenyl Column  
4mL/min  
10s 2% methanol hold  
120s 2-60% methanol gradient  
50s 60-2% methanol gradient



# Sample One Comparison: NO2 Versus EP

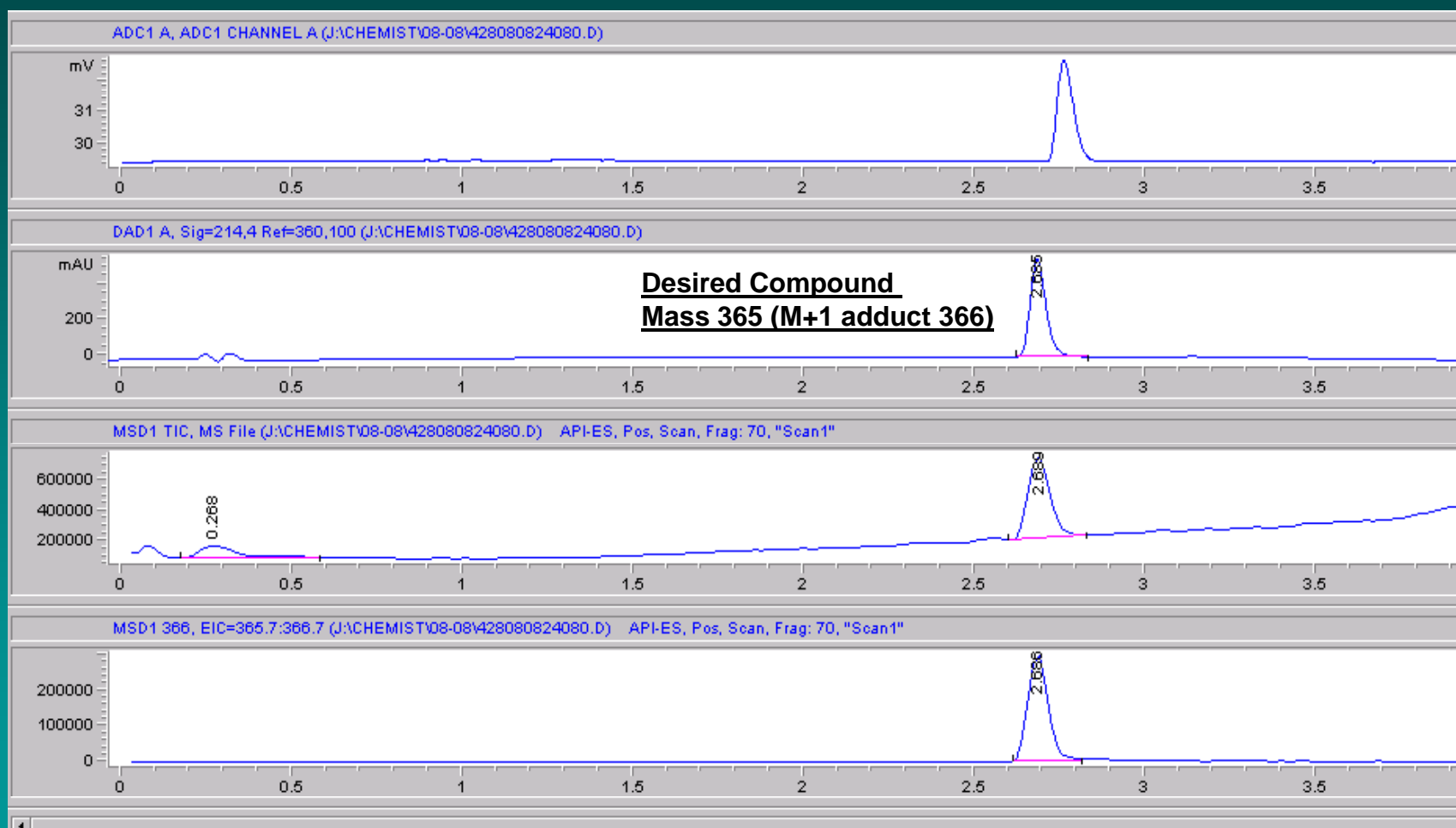


1cm i.d. x 10cm, 5 $\mu$ m, Nitrophenyl  
20mL/min, 120 Bar, 40 degrees C

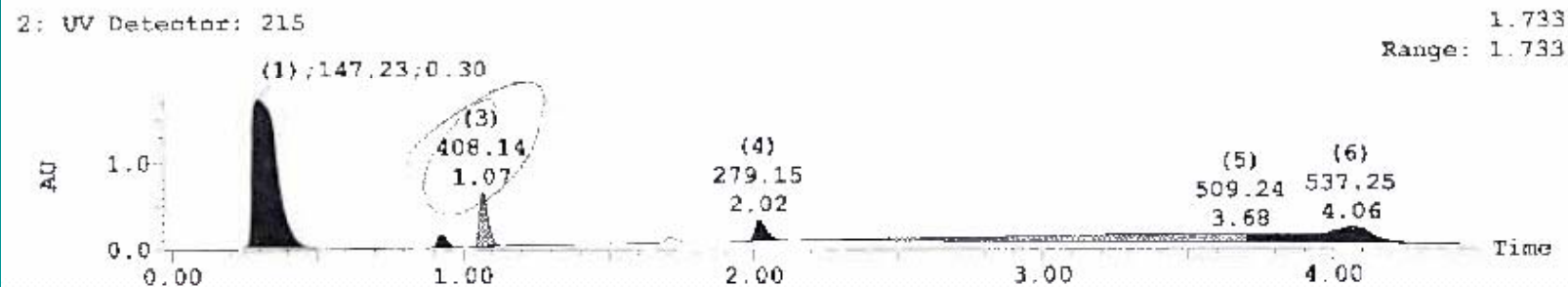
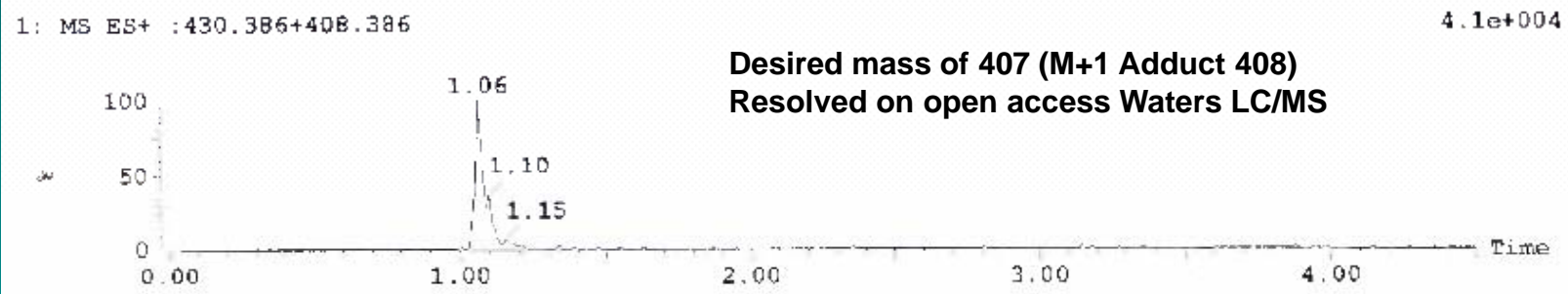
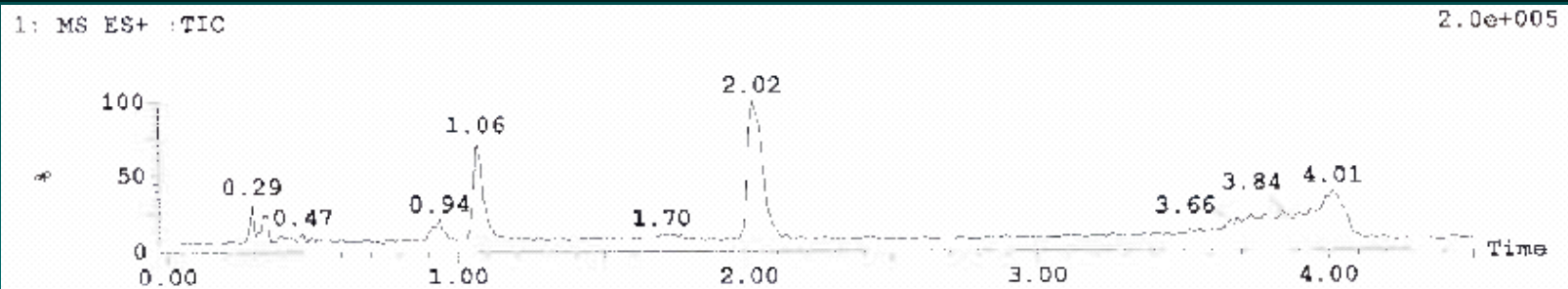
6 min 15-20% methanol  
1 min 60% flush  
2 min 60-2% reverse gradient



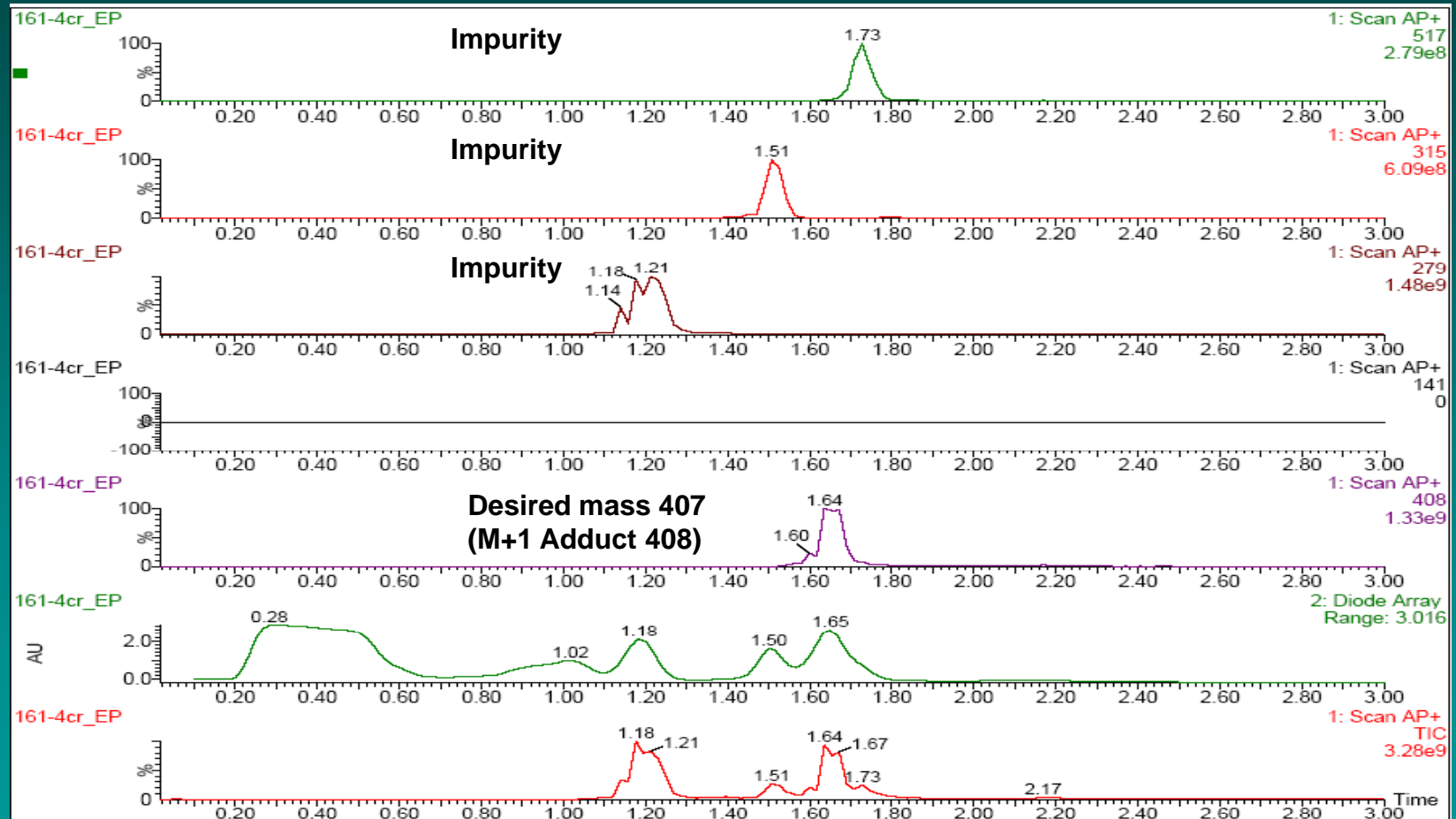
# Sample One Comparison: NO2 Versus EP



# Final Sample



# Final Sample



50mm x 4.6mm, 5um, Ethylpyridine Column

4mL/min

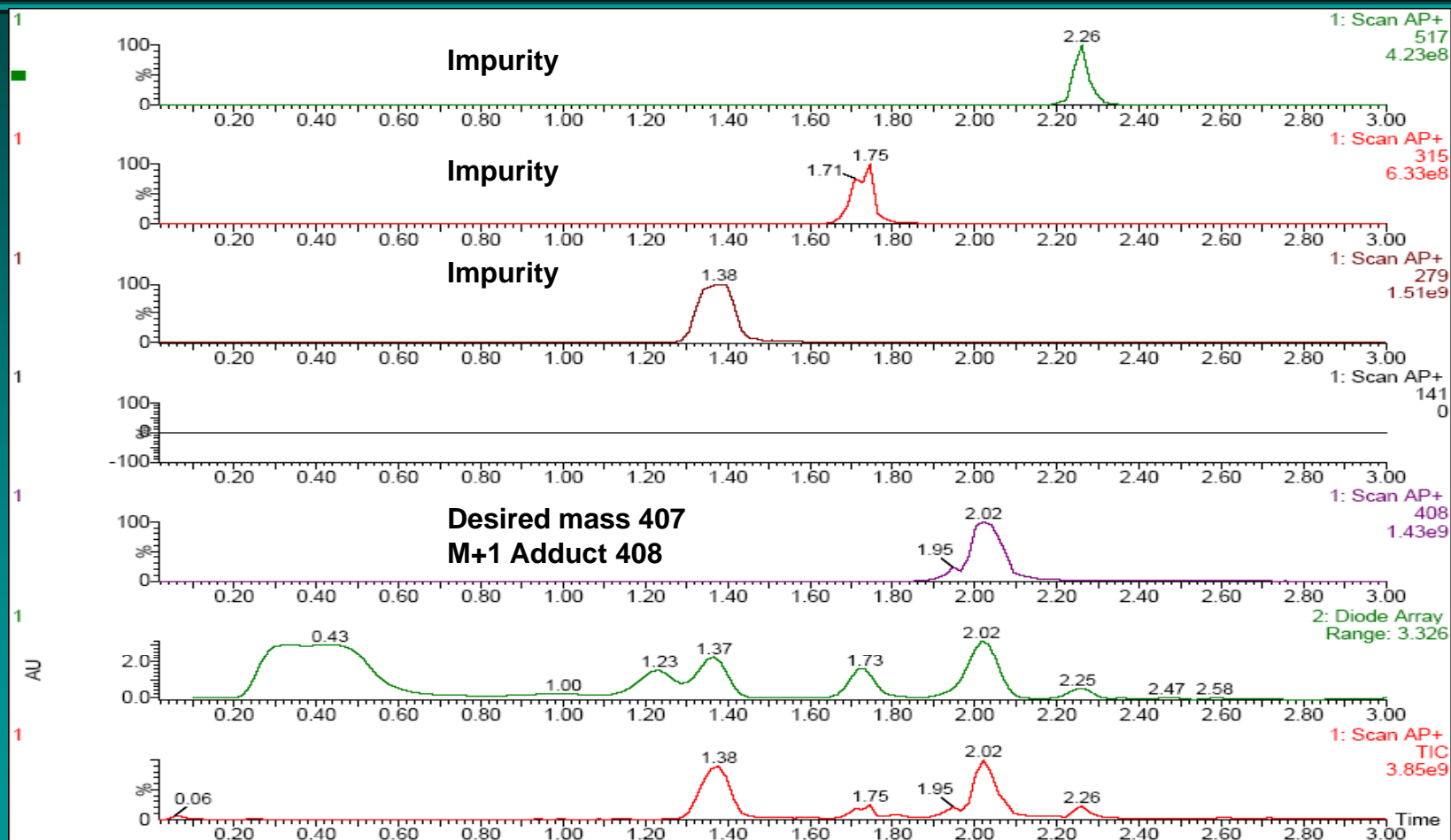
10s 2% methanol hold

120s 2-60% methanol gradient

50s 60-2% methanol gradient



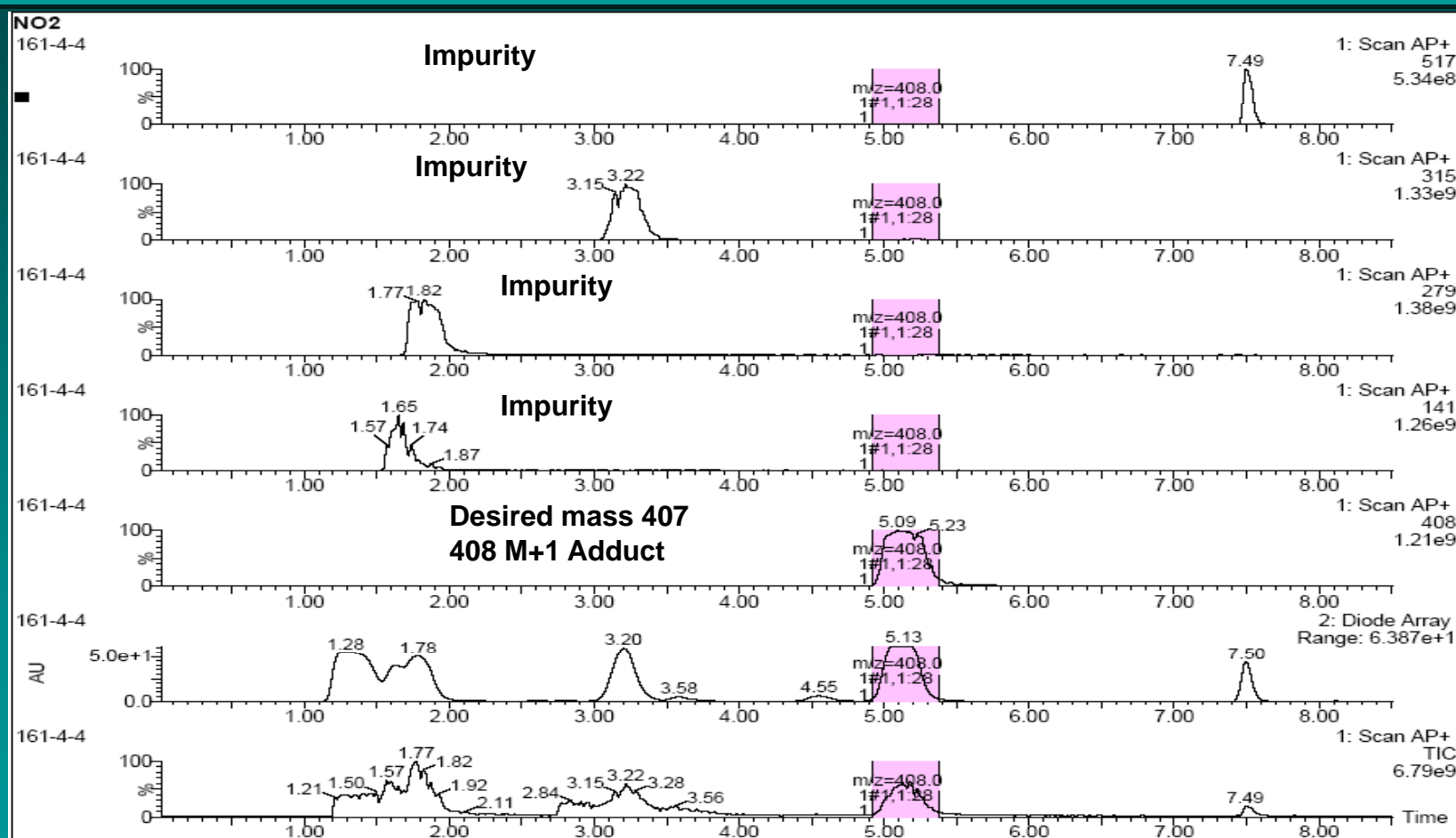
# Final Sample



50mm x 4.6mm, 5um, Nitrophenyl Column  
4mL/min  
10s 2% methanol hold  
120s 2-60% methanol gradient  
50s 60-2% methanol gradient



# Final Sample

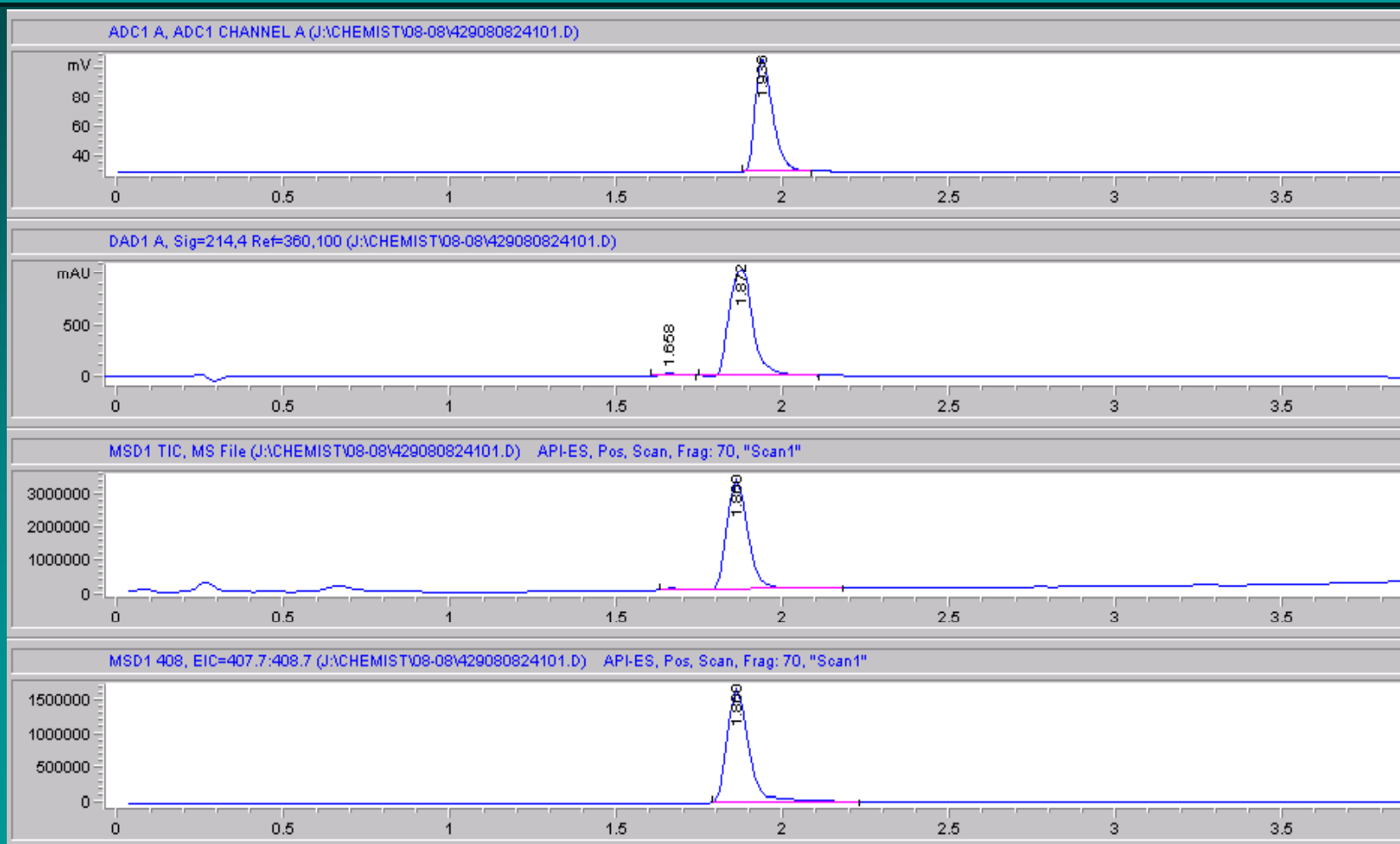


1cm i.d. x 10cm, 5 $\mu$ m, Nitrophenyl  
20mL/min, 120 Bar, 40 degrees C

6 min 20-25% methanol  
1 min 60% flush  
2 min 60-2% reverse gradient



# Final Sample



# Summary

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- Semi-preparative SFC dramatically reduces dry down time of purified compounds compared to the same scale separation on a semi-preparative HPLC system
- Analytical retention times can be mapped to focused gradients on semi-preparative SFC system for automated method assignment
- SFC is a valuable addition to a high-throughput purification lab being able to handle 60% of compounds



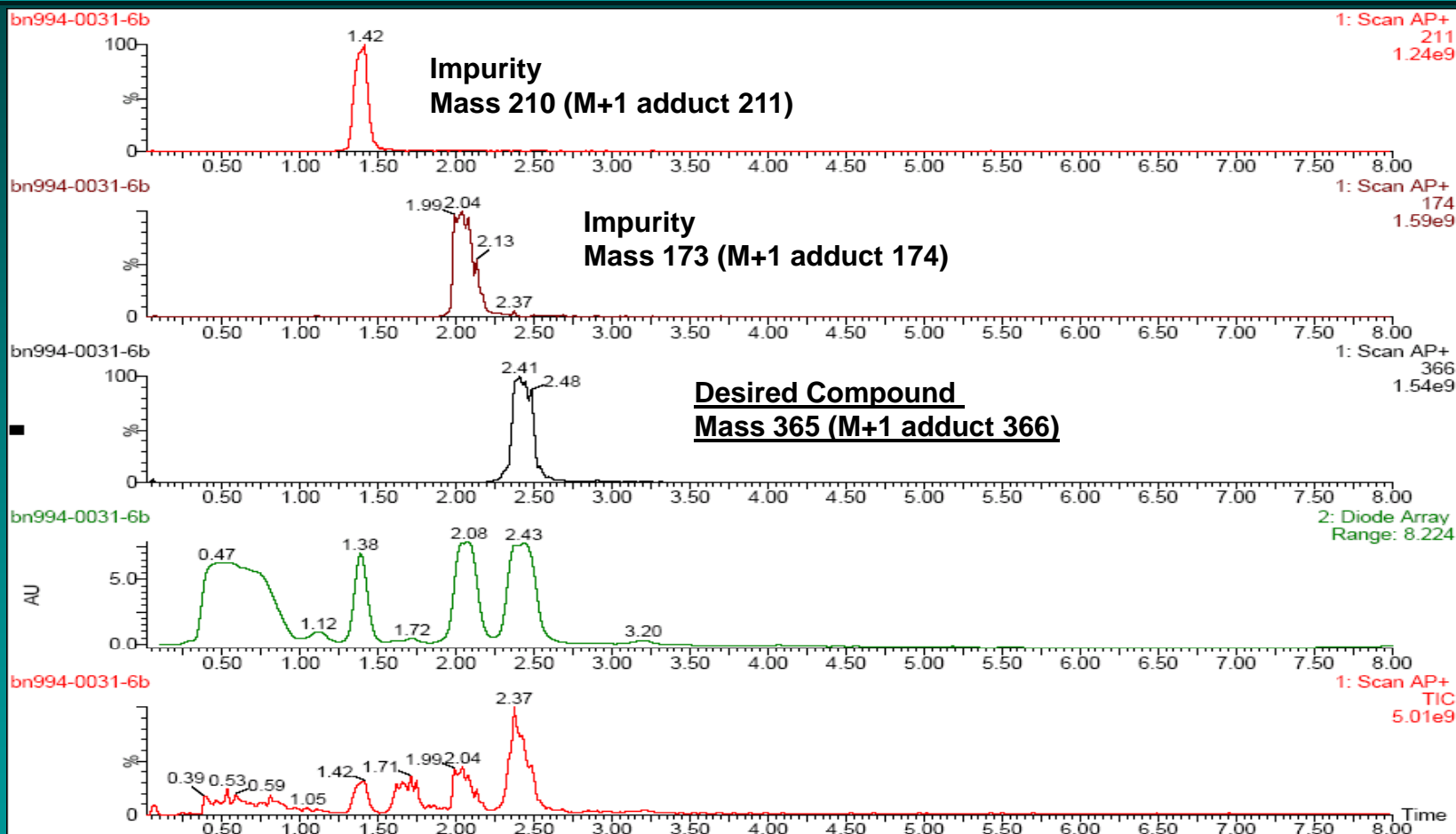
# Acknowledgements

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- Merck NTRLC
- Chris Welch, Jim Barrow, and George Hartman
- THAR
- Waters
- ES Industries
- Princeton Chromatography
- Bill Wranitz
- Matt Przybyciel
- Denise Heyburn
- And...You



# Sample One Comparison: NO2 Versus EP



50mm x 4.6mm, 5um, Ethylpyridine Column

4mL/min

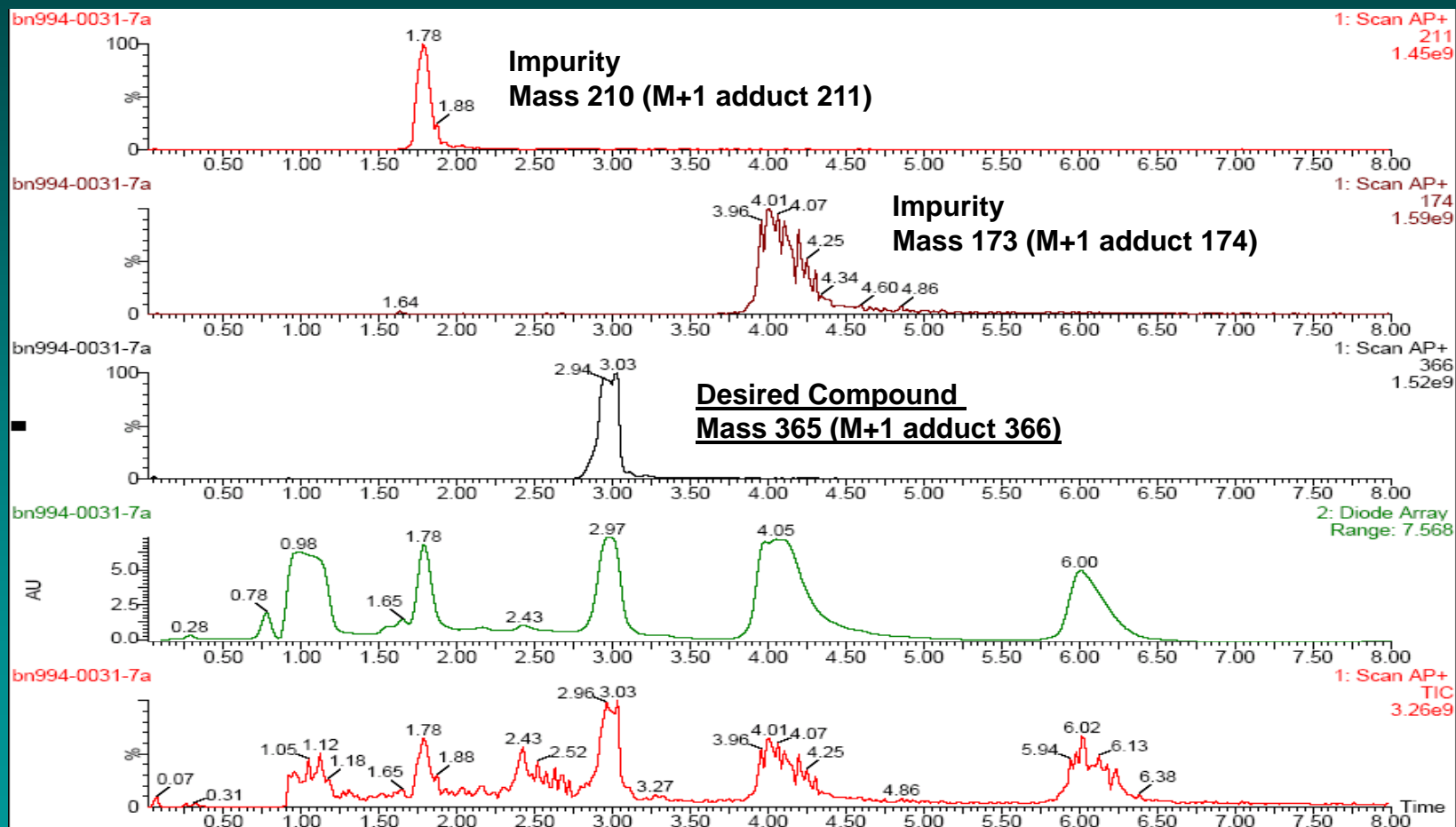
10s 2% methanol hold

360s 2-60% methanol gradient

110s 60-2% methanol gradient



# Sample One Comparison: NO2 Versus EP



50mm x 4.6mm, 5um, Nitrophenyl Column

4mL/min

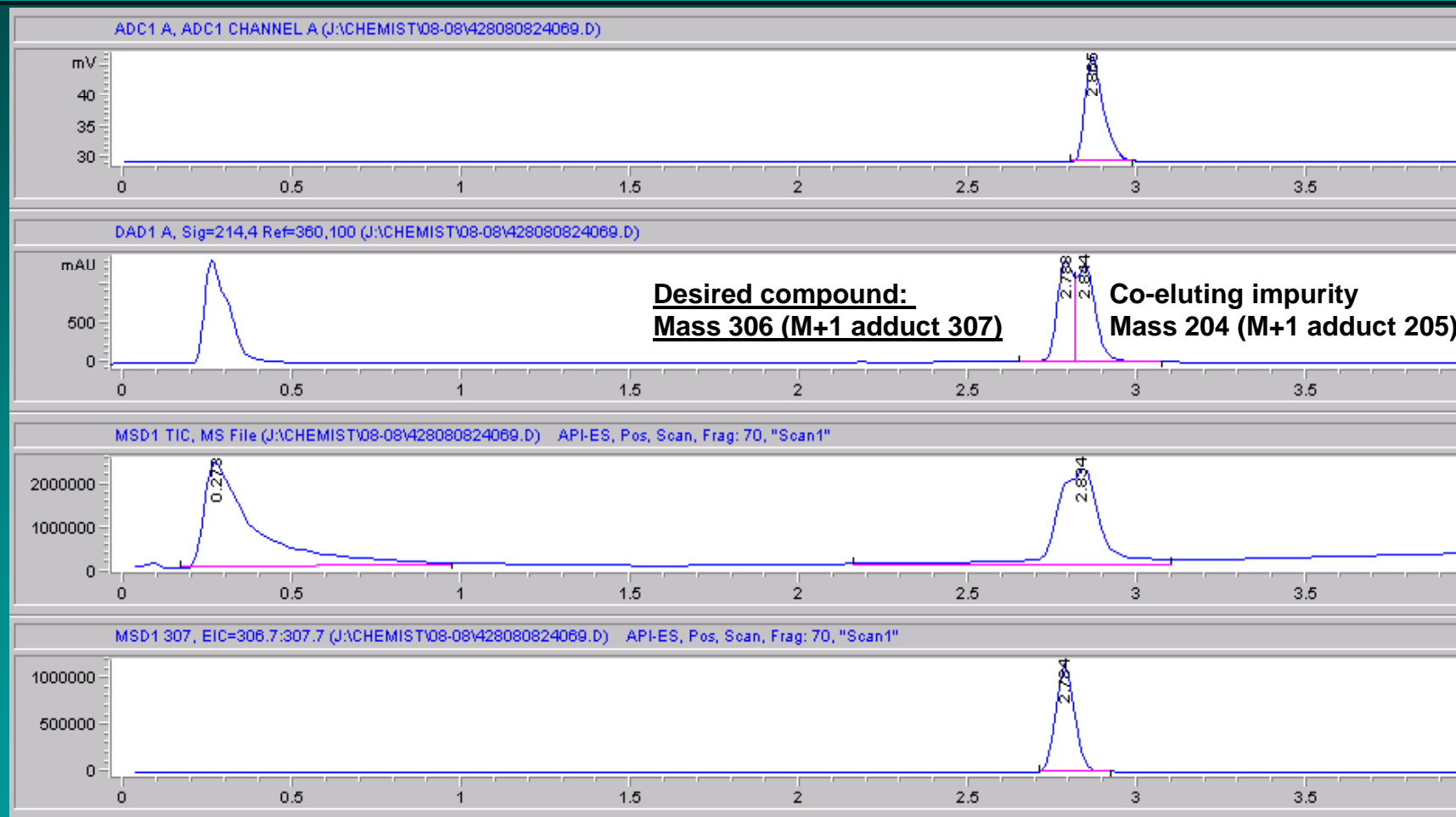
10s 2% methanol hold

360s 2-60% methanol gradient

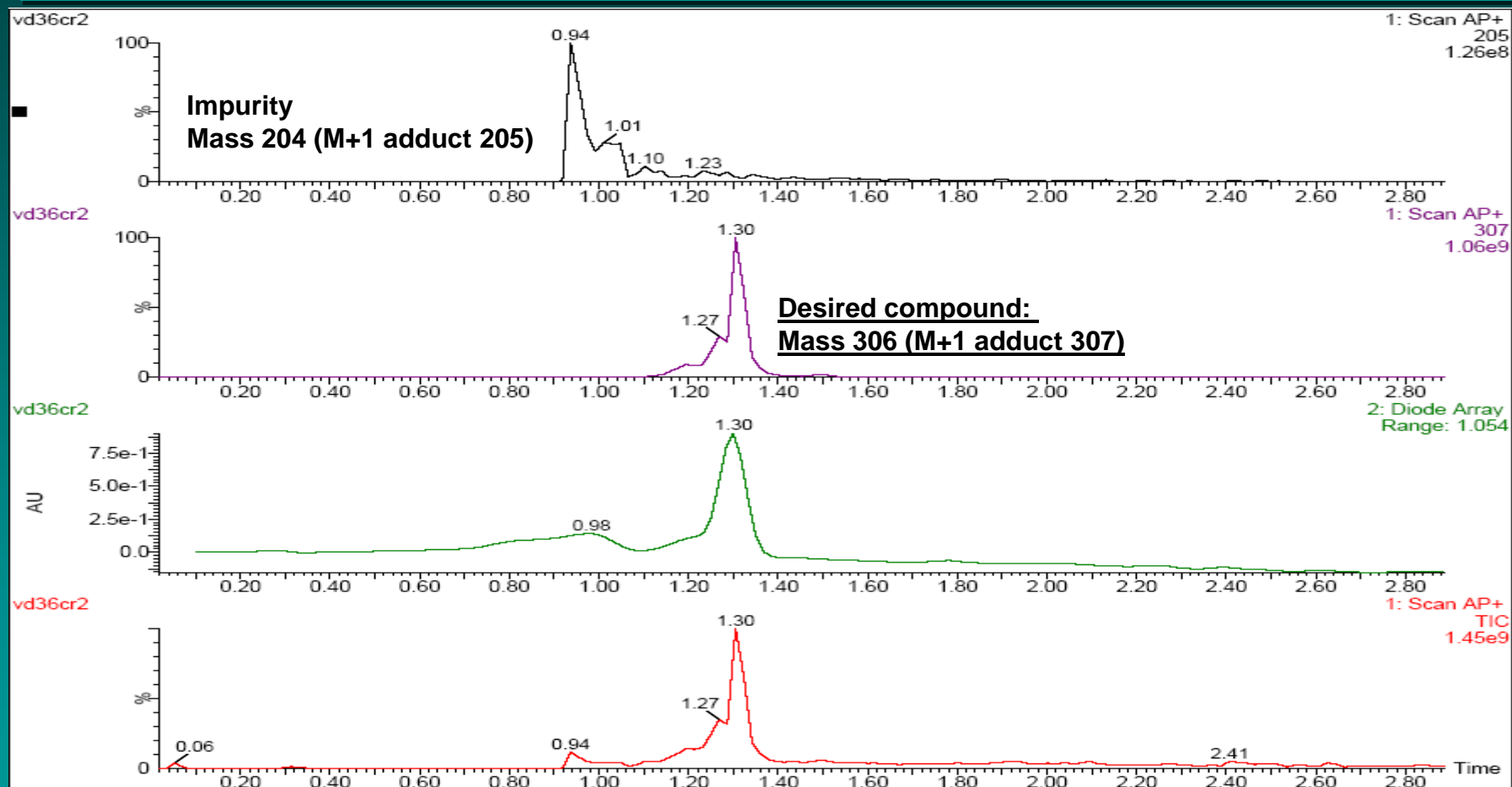
110s 60-2% methanol gradient



# Sample Two Comparison: NO<sub>2</sub> Versus EP



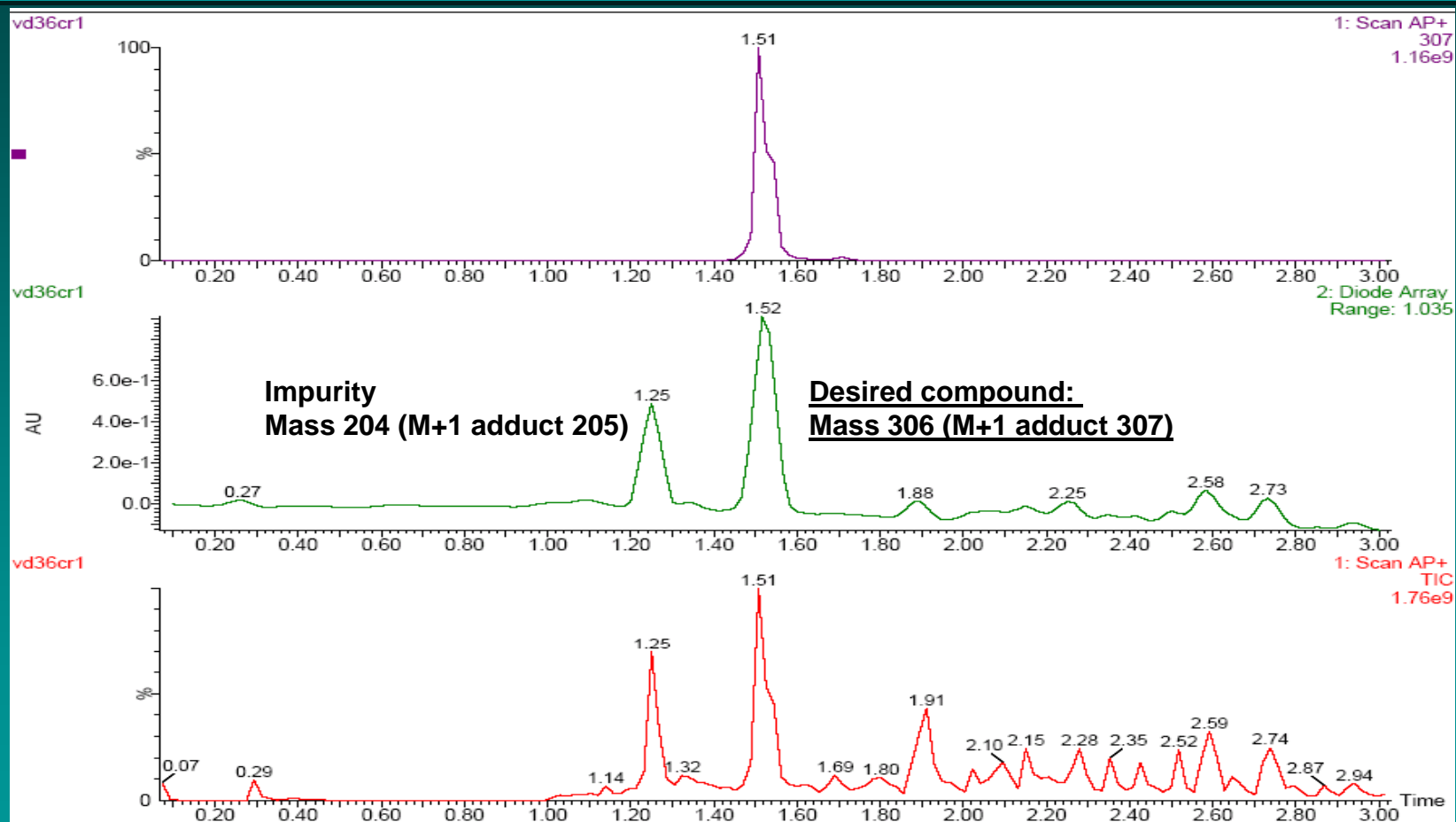
# Sample Two Comparison: NO<sub>2</sub> Versus EP



50mm x 4.6mm, 5um, Ethylpyridine Column  
4mL/min  
10s 2% methanol hold  
120s 2-60% methanol gradient  
50s 60-2% methanol gradient



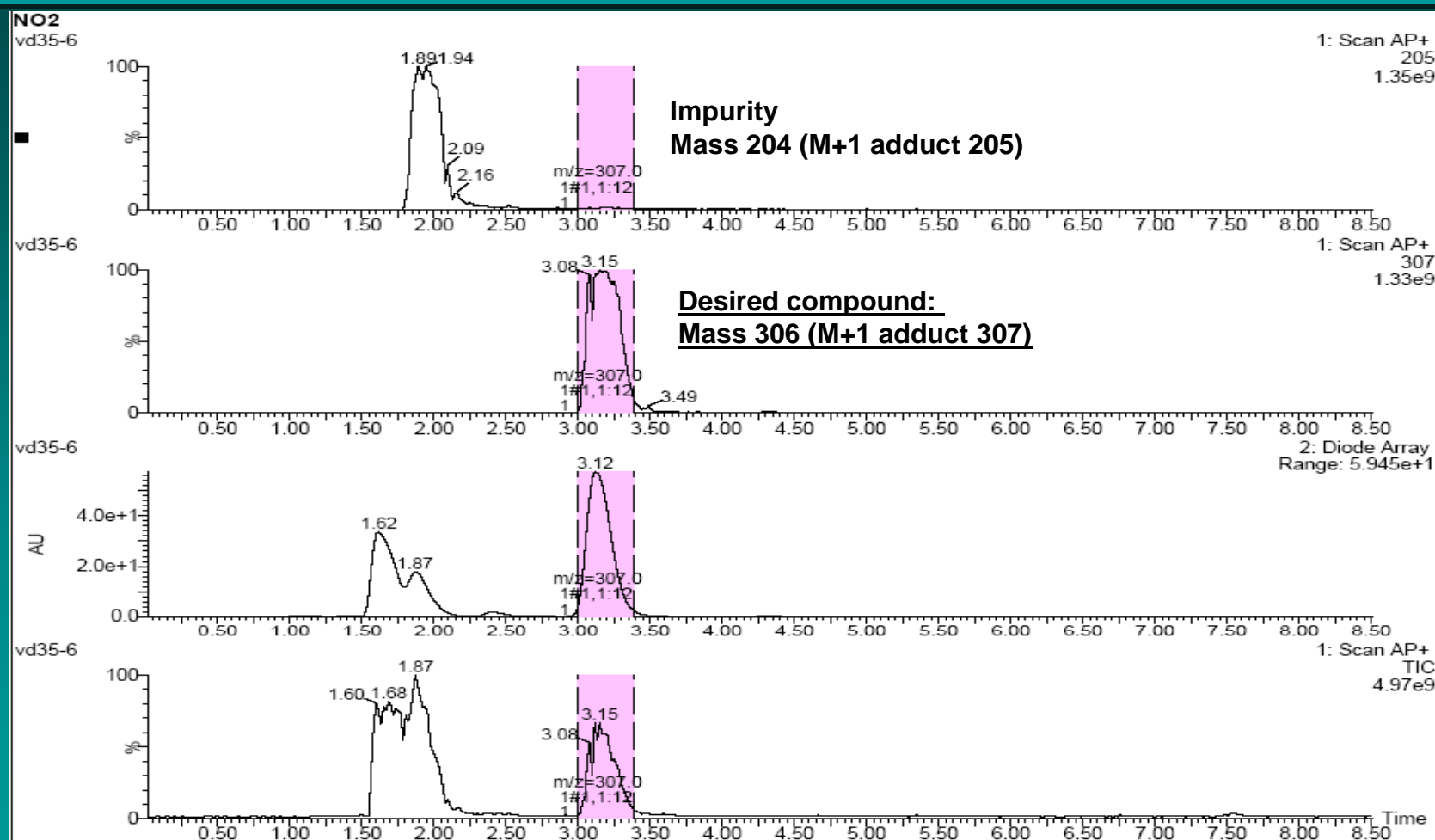
# Sample Two Comparison: NO2 Versus EP



50mm x 4.6mm, 5um, NitrophenylColumn  
4mL/min  
10s 2% methanol hold  
120s 2-60% methanol gradient  
50s 60-2% methanol gradient



# Sample Two Comparison: NO2 Versus EP



1cm i.d. x 10cm, 5 $\mu$ m, Nitrophenyl  
20mL/min, 120 Bar, 40 degrees C

6 min 15-20% methanol  
1 min 60% flush  
2 min 60-2% reverse gradient



# Sample Two Comparison: NO2 Versus EP

